



Unicycler: bacterial genome assemblies from short and long read sequences

Journal Club

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RESEARCH ARTICLE

Unicycler: Resolving bacterial genome assemblies from short and long sequencing reads

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<https://github.com/rrwick/Unicycler#2022-update>

What is Unicycler

= Assembly pipeline for bacterial genomes

Use cases:

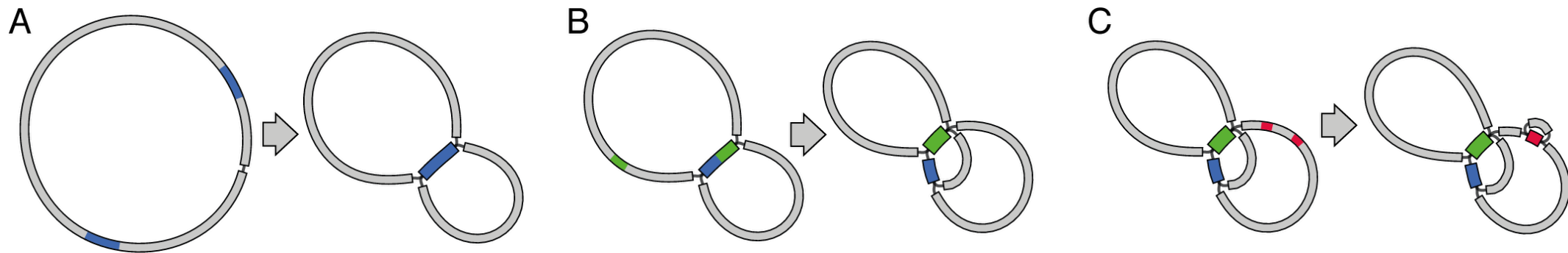
- Short reads only
- Long reads only
- **Short and long reads** from the same isolate (best case)

Why use Unicycler:

- It circularizes replicons
- Produces assembly graph in addition to contigs FASTA file
- Handles plasmid-rich or repetitive genomes
- Can use long reads of any depth and quality
- Filters out low-depth contigs (useful in case of low level contamination)

Limitations of short reads

- Accurate but short reads are smaller than many repetitive elements in bacterial genomes
- Assembly of a full genome often not possible
- Incomplete genomes hinder large-scale comparative genomic studies
- **Short** reads don't have enough info to resolve repeats but **long** reads do => **hybrid assembly**



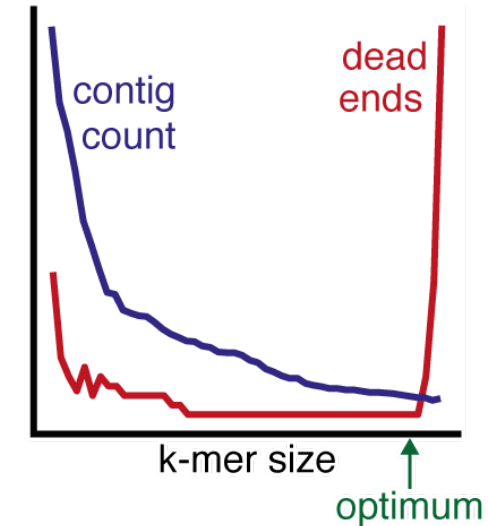
As repeats are added, the graph becomes increasingly tangled

Source: <https://github.com/rwick/Unicycler#2022-update>

Unicycler pipeline

SPAdes

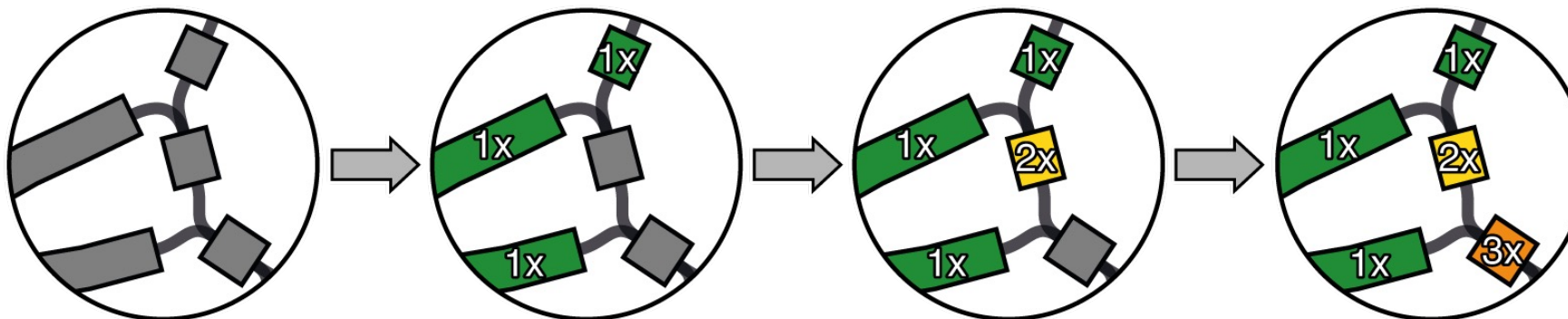
- Short read assembly
- Made by performing a **De Bruijn graph** assembly with a **wide range of k-mer** size
- Each assembly builds on the previous one, allowing SPAdes to get the advantages from:
 - Small k-mer assemblies = more connected graph
 - Large k-mer assemblies = better resolved repeats
- When assembling Illumina reads,
 - Unicycler** functions as SPAdes optimizer



Unicycler pipeline

Multiplicity

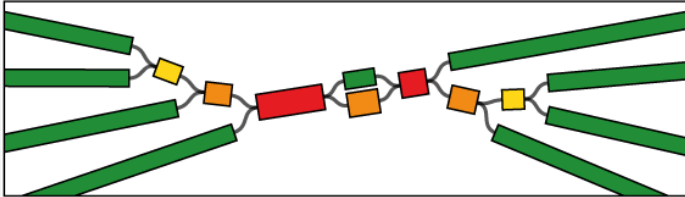
- Goal: distinguish between single copy contigs and repeat contigs
- Read depth can be indicative of multiplicity if genome single chromosome
- Greedy algorithm uses **read depth** and **connectivity**
- Multiplicity of 1 assigned to contigs close to median depth and with one connection at either end



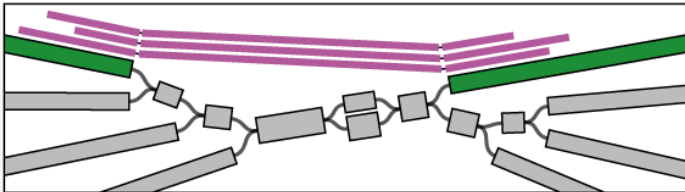
Bridging

- Scaffolding by building bridges between single copy contigs using the path information in the SPAdes assembly
- Bridges made using long reads can resolve larger repeats

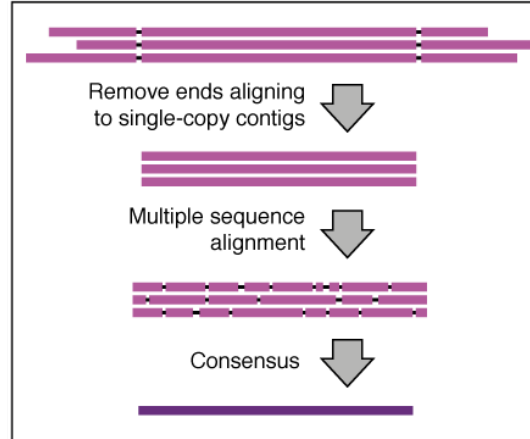
Repeat region in unbridged graph



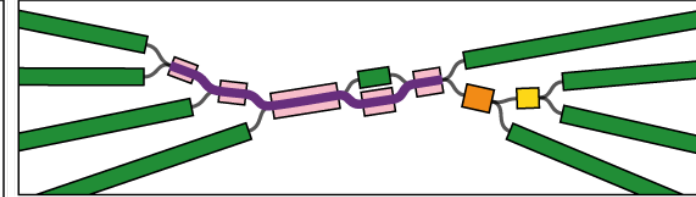
Semi-global long read alignment



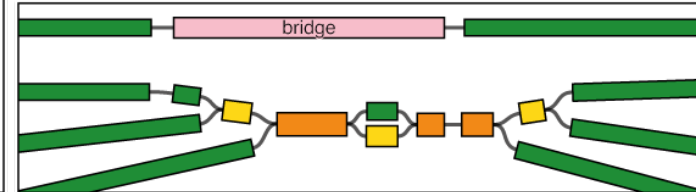
Consensus read sequence



Path finding



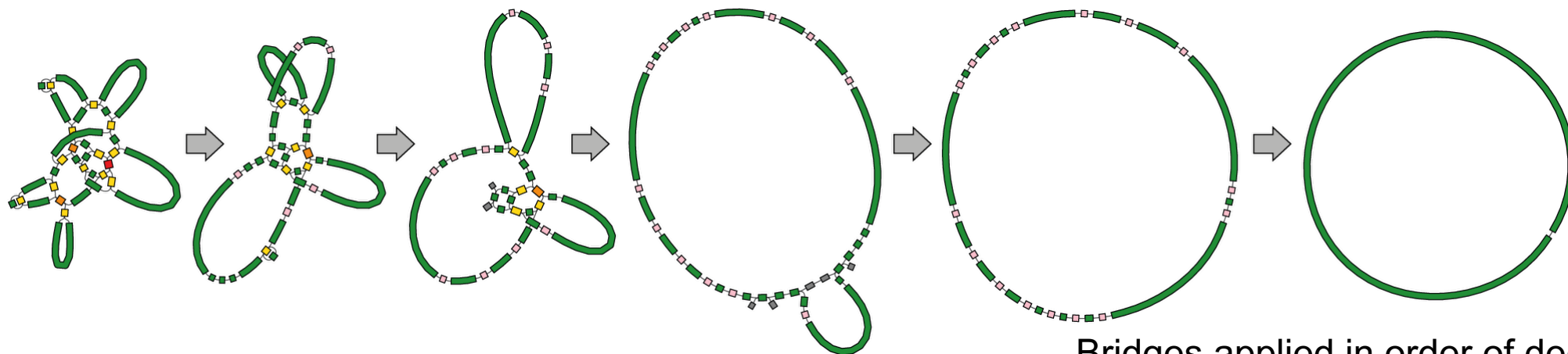
Bridged graph



Unicycler pipeline

Bridge application

- Multiple possible bridges, some may be conflicting
- **Quality score** assigned to each bridge
 - Number of reads supporting the bridge
 - Length of contigs to be bridged
 - Alignment quality between read consensus and graph path
 - Read depth consistency between contigs

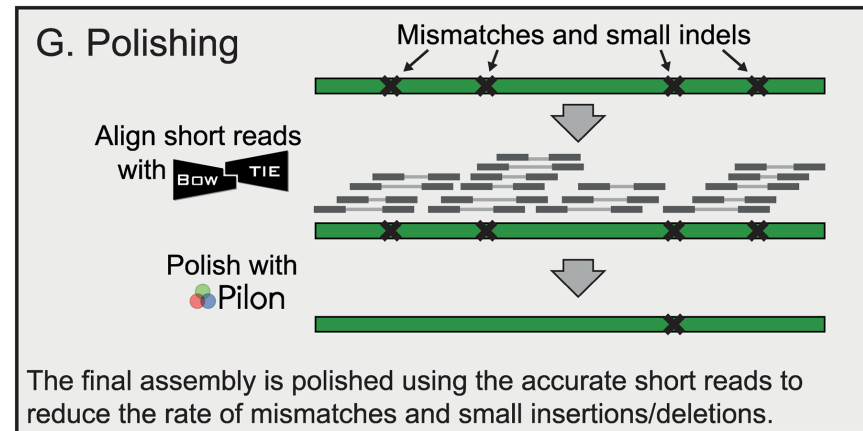
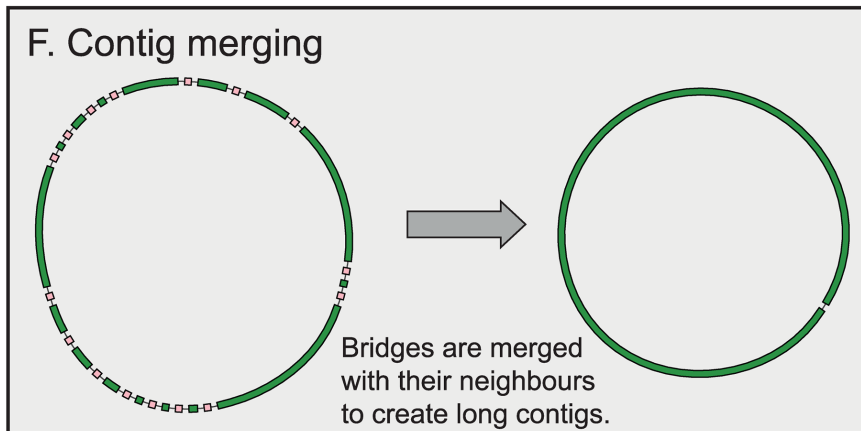


Bridges applied in order of decreasing quality

Unicycler pipeline

Merging & polishing

- Reducing rate of mismatches and small errors using the accurate short reads
- Pilon



Modes

Conservative

- Bridge quality cutoff high
- Low risk of misassembly
- Least likely to produce a complete genome
- Use when high accuracy needed


Normal

- Bridge quality cutoff intermediate

Bold

- Lower quality bridges used
- Greater risk of error
- Use when completeness more important than accuracy

Performance evaluation of Unicycler

1. **Simulated read** sets from 12 **reference** genomes
 2. **Real** read sets from *E.coli* K-12 
 3. **Novel** *Klebsiella pneumoniae* isolate genome (no reference genome)
- Comparison to other pipelines and tools

Misassemblies

- cases where a contig aligns to the reference genome in multiple pieces, not as a single continuous alignment, indicating a structural error in the contig
- For simulated reads, misassemblies indicate assembler mistake
- In *E.coli*, false positive misassemblies also possible

Small-error rate

- Mismatches
- Indels

Quast

NGA50

- Length of contig-to-reference alignment
- A correctly assembled contig will have a single, unbroken alignment to the reference
- A misassembled contig will be divided into multiple smaller alignments

Simulated read sets

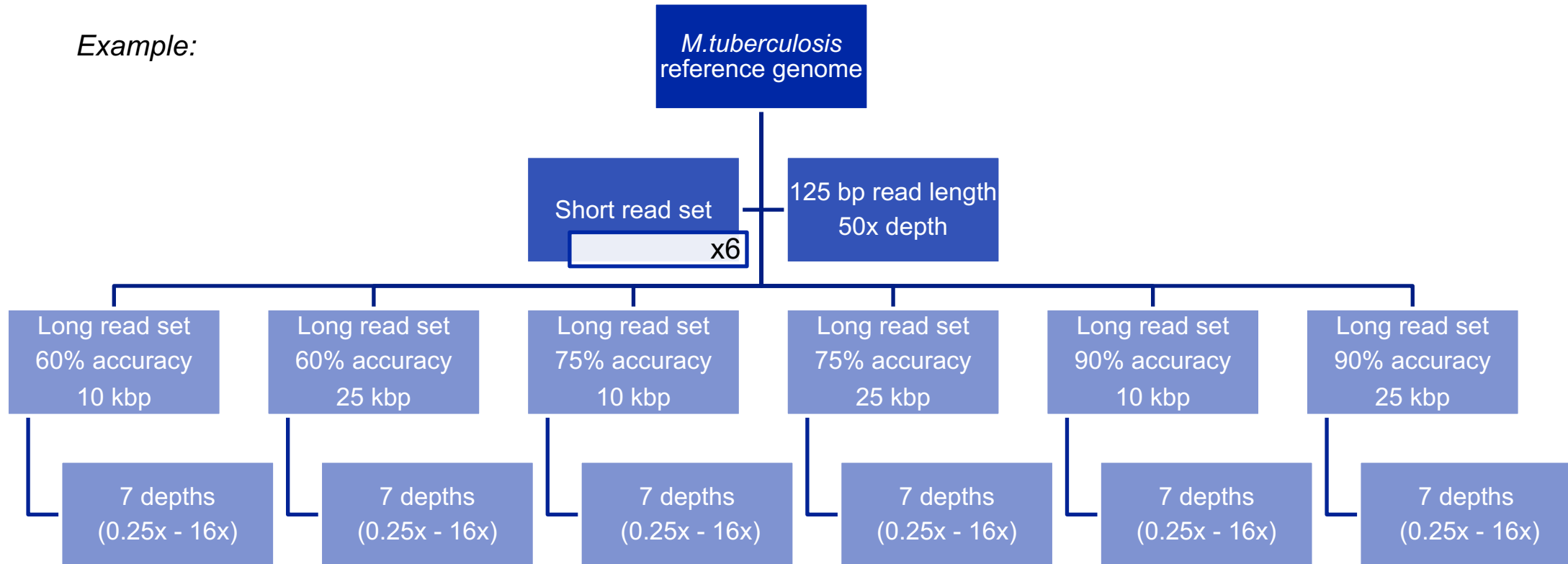
Species	Strain	Genome size (bp)	GC content	Description	MLST sequence type	Other features	GenBank assembly accession
<i>Acinetobacter baumannii</i>	A1	3,917,739	39.3%	Circular chromosome, one plasmid	231	Large, repetitive biofilm-associated protein gene	GCA_000830055.1
<i>Acinetobacter baumannii</i>	AB30	4,335,793	39.0%	Circular chromosome	758	Large, repetitive biofilm-associated protein gene	GCA_000746645.1
<i>Escherichia coli</i>	K-12 MG1655	4,641,652	50.8%	Circular chromosome	10		GCA_000005845.2
<i>Escherichia coli</i>	O25b: H4-ST131 EC958	5,249,449	50.8%	Circular chromosome, two plasmids	131		GCA_000285655.3
<i>Klebsiella pneumoniae</i>	30660/ NJST258_1	5,540,936	57.2%	Circular chromosome, five plasmids	258		GCA_000598005.1
<i>Klebsiella pneumoniae</i>	MGH 78578	5,694,894	57.1%	Circular chromosome, five plasmids	38		GCA_000016305.1
<i>Klebsiella pneumoniae</i>	NTUH-K2044	5,472,672	57.4%	Circular chromosome, one plasmid	23		GCA_000009885.1
<i>Mycobacterium tuberculosis</i>	H37Rv	4,411,532	65.6%	Circular chromosome		High-copy-number PE and PPE genes	GCA_000195955.2
<i>Saccharomyces cerevisiae</i>	S288c	12,157,105	38.1%	16 linear chromosomes, circular mitochondrial sequence		Eukaryote	GCA_000146045.2
<i>Shigella dysenteriae</i>	Sd197	4,560,911	51.0%	Circular chromosome, two plasmids	146	High insertion sequence content	GCA_000012005.1
<i>Shigella sonnei</i>	53G	5,220,473	50.7%	Circular chromosome, four plasmids	152	High insertion sequence content	GCA_000283715.1
<i>Streptococcus suis</i>	BM407	2,170,808	41.0%	Circular chromosome, one plasmid	1		GCA_000026745.1

- Variety of genome sizes and complexities

<https://doi.org/10.1371/journal.pcbi.1005595.t001>

Simulated read sets

Example:



- For each strain there are 6 short-read sets and 42 hybrid-read sets

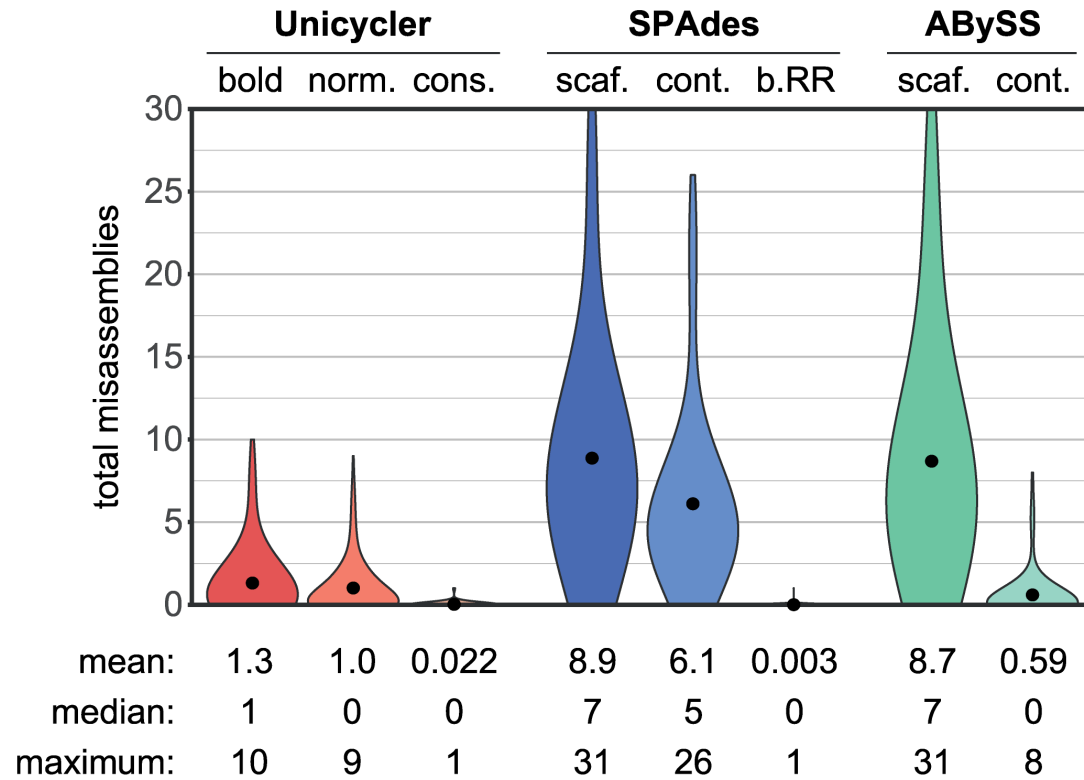
Comparing to different pipelines/tools

Short read assembly evaluation Hybrid assembly evaluation

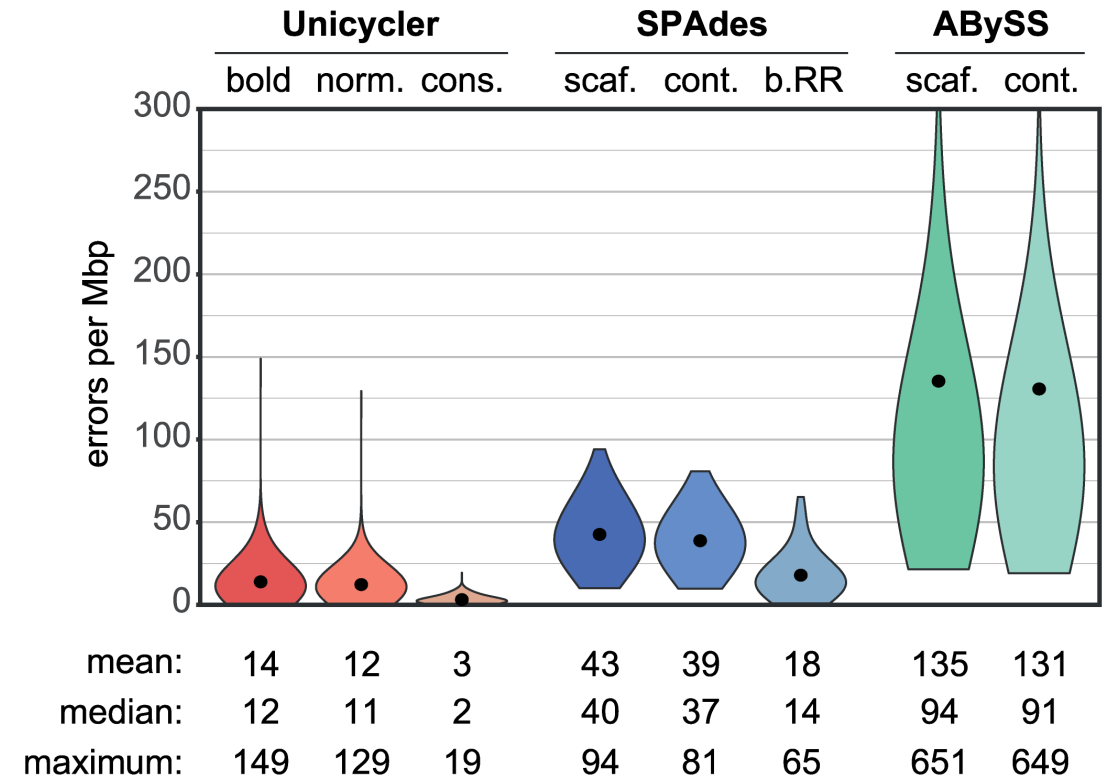
- Unicycler
 - SPAdes
 - ABySS
- Unicycler
 - SPAdes
 - npScarf
 - Cerulean

Simulated read sets – short read assembly evaluation

Misassemblies



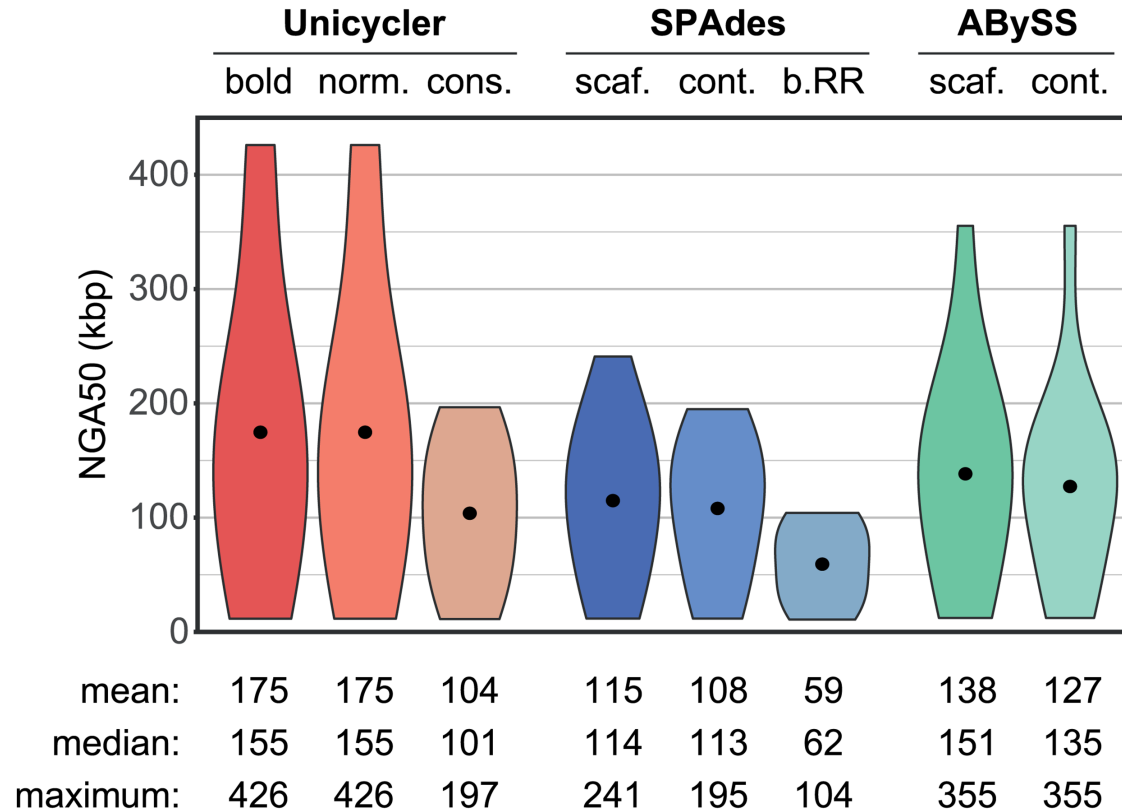
Small errors



n=360 per assembler

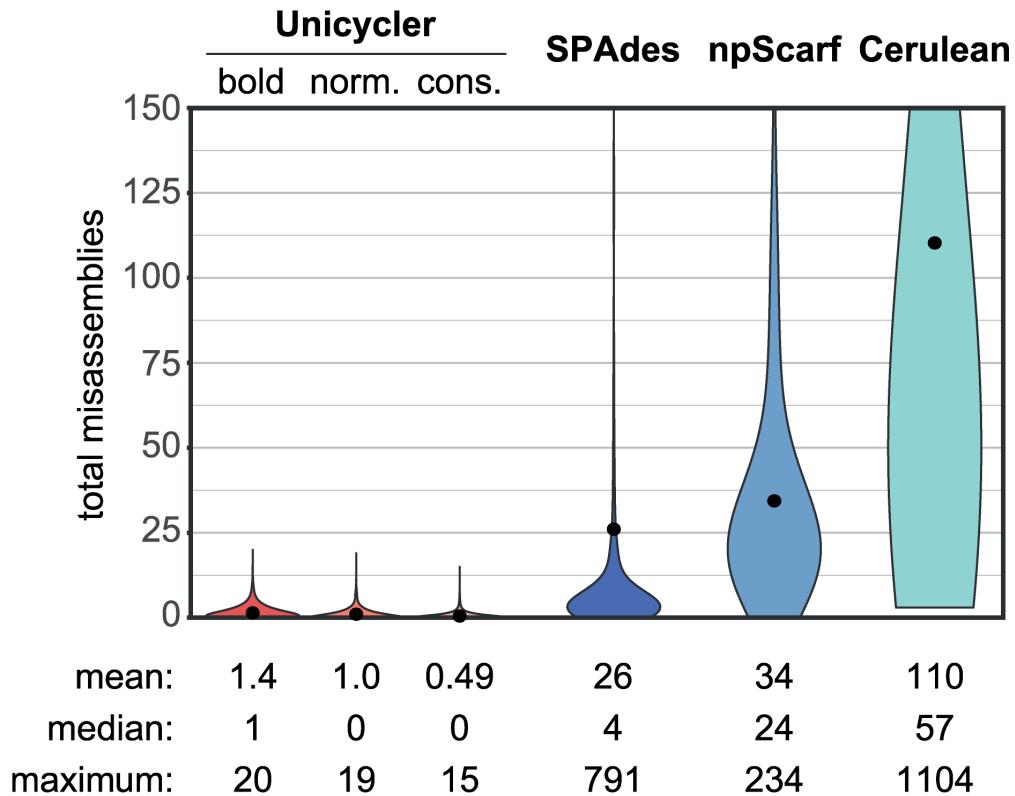
Simulated read sets – short read assembly evaluation

NGA50

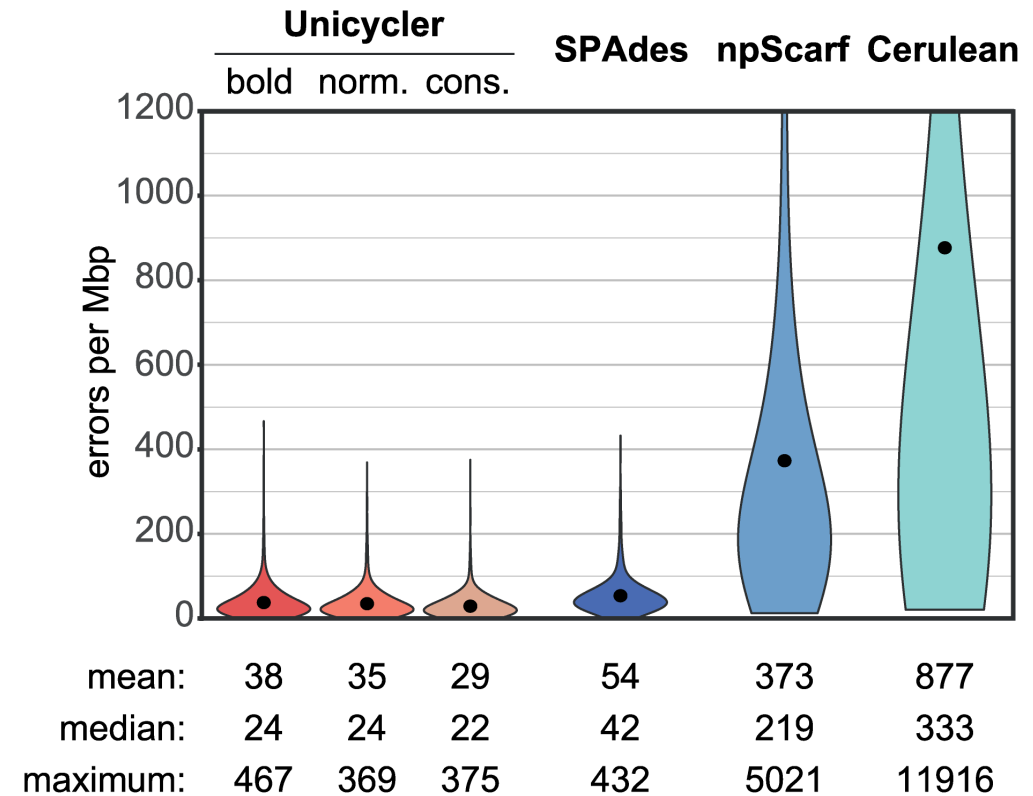


Simulated read sets – hybrid assembly evaluation

Misassemblies



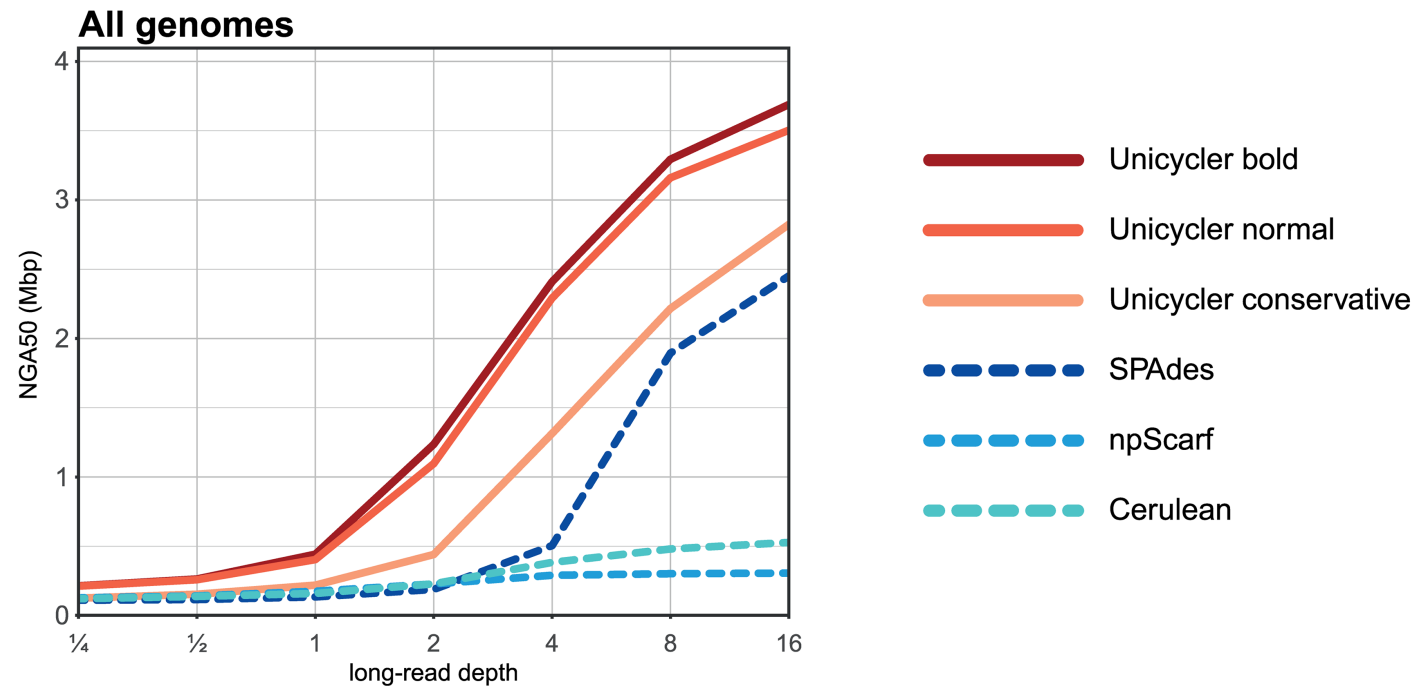
Small errors



n=2520 per assembler

Simulated read sets – hybrid assembly evaluation

NGA50

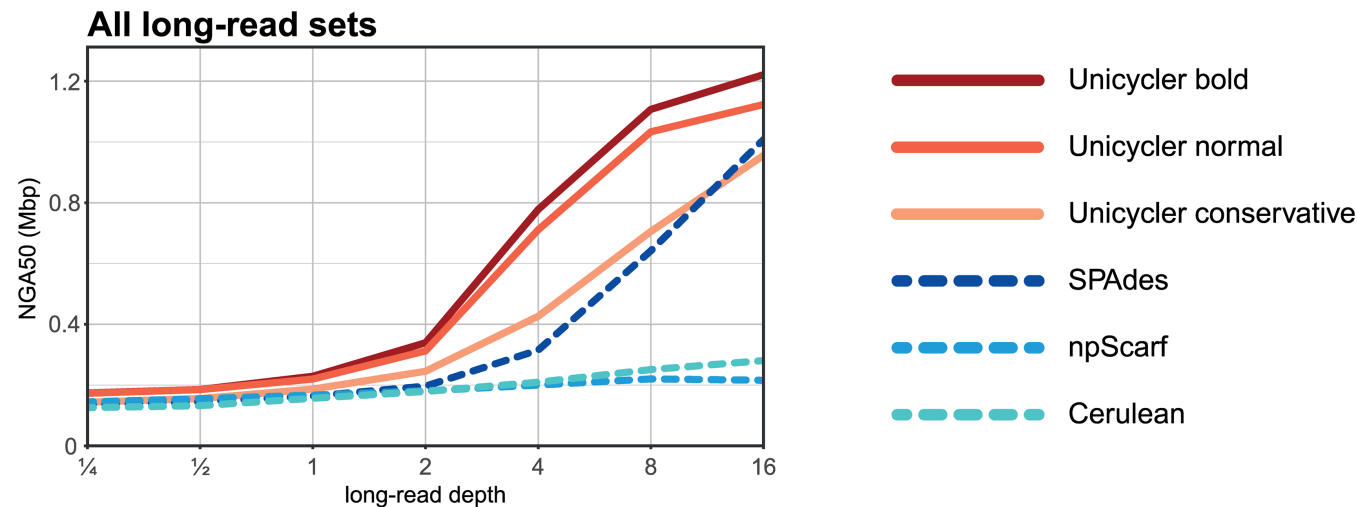


Real *E.coli* K-12 read sets – hybrid assembly

- Short reads produced on Illumina MiSeq
- Long reads from different platforms (ONT, PacBio, different flow cells and chemistries)
- Accuracy assessed by comparison to the reference genome (Sanger-based capillary sequencing at the University of Wisconsin in 1997)

Unicycler produced larger contigs at lower long-read depths than other assemblers

NGA50



NGA50 overall lower compared to simulated reads

Why?

Klebsiella pneumoniae de-novo assembly

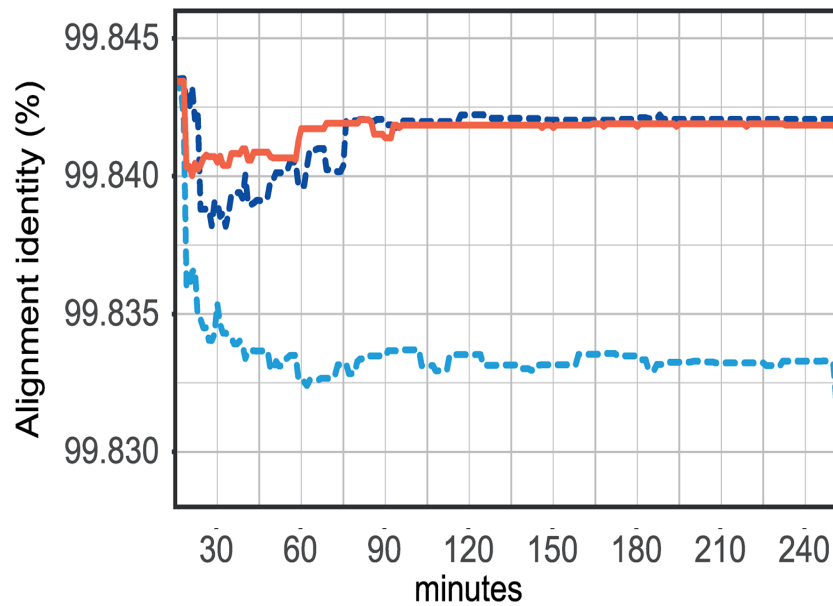
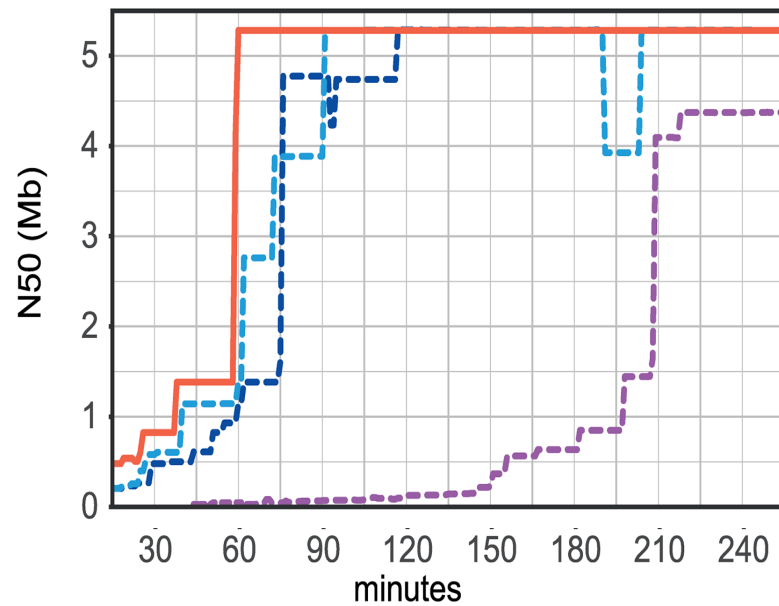
- *K.pneumoniae* isolate INF125
- Virulent strain from urine of a Melbourne hospital patient
- **Real time analysis**
 - Sequencing can be stopped when complete assembly reached

Results:

Overall time?

- 240 subsets of reads (after each minute of sequencing)
 - **Unicycler** completed assebbly with data generated in 45 min (depth = 5.3x)
 - **npScarf** with data generated in 76 min (9x)
 - **SPAdes** with data generated in 102 min (12.1x)
 - **Miniasm** with data generated in 213 min (25.3x)

Klebsiella pneumoniae de-novo assembly



- Unicycler normal
- SPAdes
- npScarf
- miniasm

Summary

- Unicycler performed well on both short-read-only and hybrid-read sets
- Larger contigs than other assemblers
- Fewer misassemblies
- Genome sufficiently resolved with low-depth long reads (conserving resources)

- Unicycler was initially made in 2016, back when long reads were sparse and noisy
- Unicycler was designed to use low-depth and low-accuracy long reads to scaffold a short-read assembly graph to completion, i.e. short-read-first hybrid assembly
- Nowadays, Nanopore sequencing yield is now much higher and **improved accuracy**
- **Long-read-first assembly** is a viable approach (**Trycycler** and **Polypolish**)
- **Unicycler** still the best tool for **short-read-first hybrid assembly** of bacterial genomes and short-read-only bacterial genome assembly

Future

- Improved accuracy of long read sequencing
- Long read only assembly
- Real time assembly and stopping sequencing when assembly complete
- Ultimately lowered costs

Thank you

Questions?

