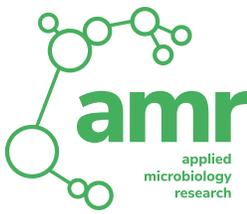




University of
Zurich^{UZH}

Institute of Medical Microbiology

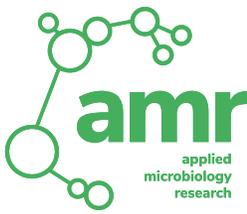


Introduction to Typing

Helena Seth-Smith PhD

23.03.2023

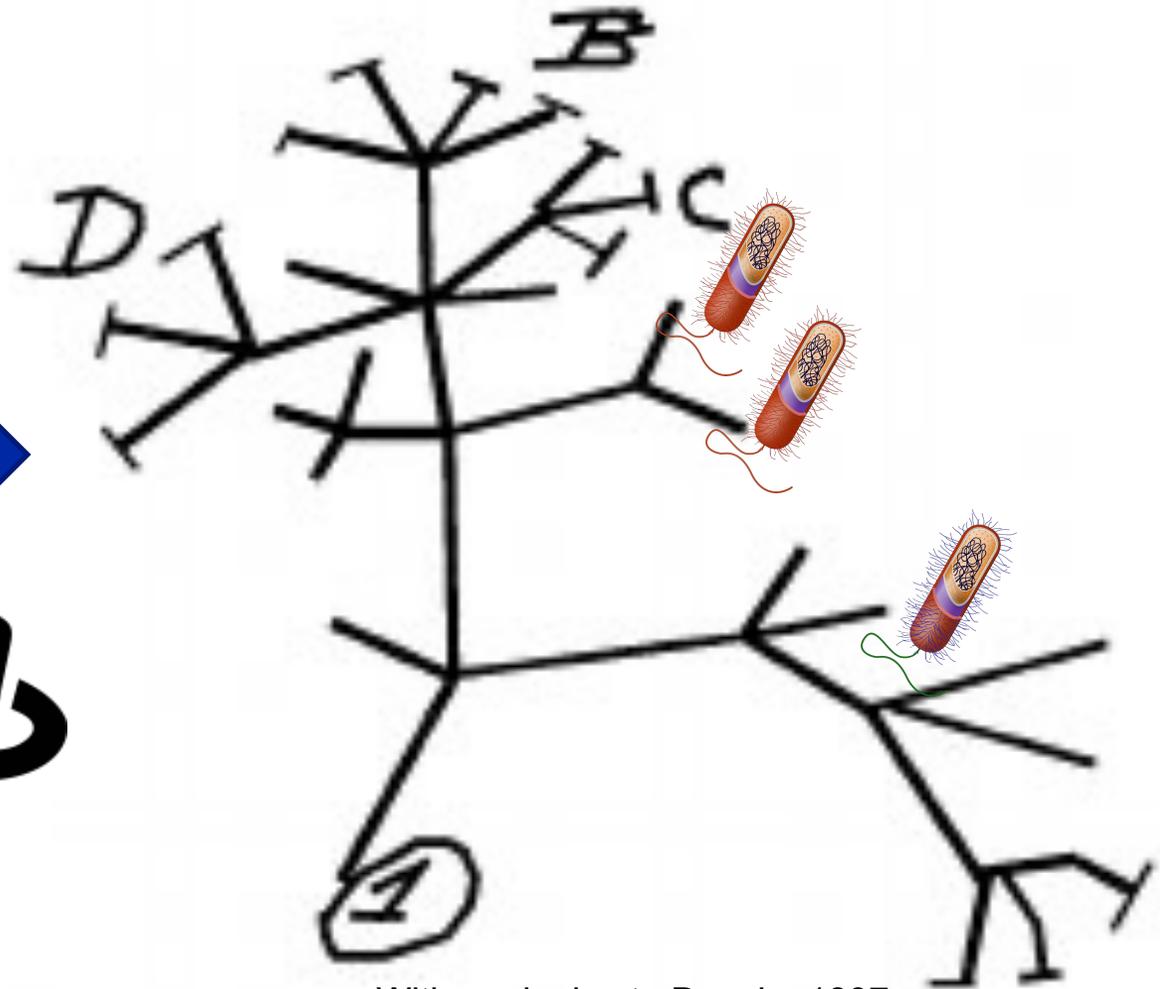
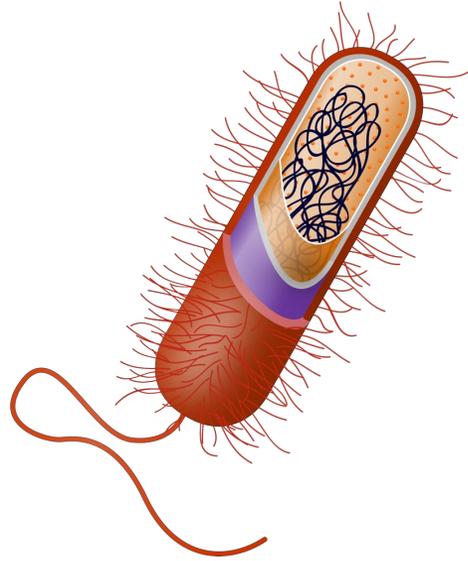
Table of Contents



- What is Typing?
- Conventional typing
- Molecular typing
- Examples
- Resolution of genome-based typing methods
- MLST
- Examples
- Recombination
- Summary

What is Typing?

A method of categorizing microbes



With apologies to Darwin, 1837

Conventional methods of typing

“Microbial typing is often employed to determine the source and routes of infections, confirm or rule out outbreaks, trace cross-transmission of healthcare-associated pathogens, recognize virulent strains and evaluate the effectiveness of control measures” (Ranjbar *New Microbiologica* 2014)

Conventional typing was phenotypic:

- Gram staining
- Serotyping: based on surface characteristics recognized by immune system (eg rabbit antisera)
- Phage typing

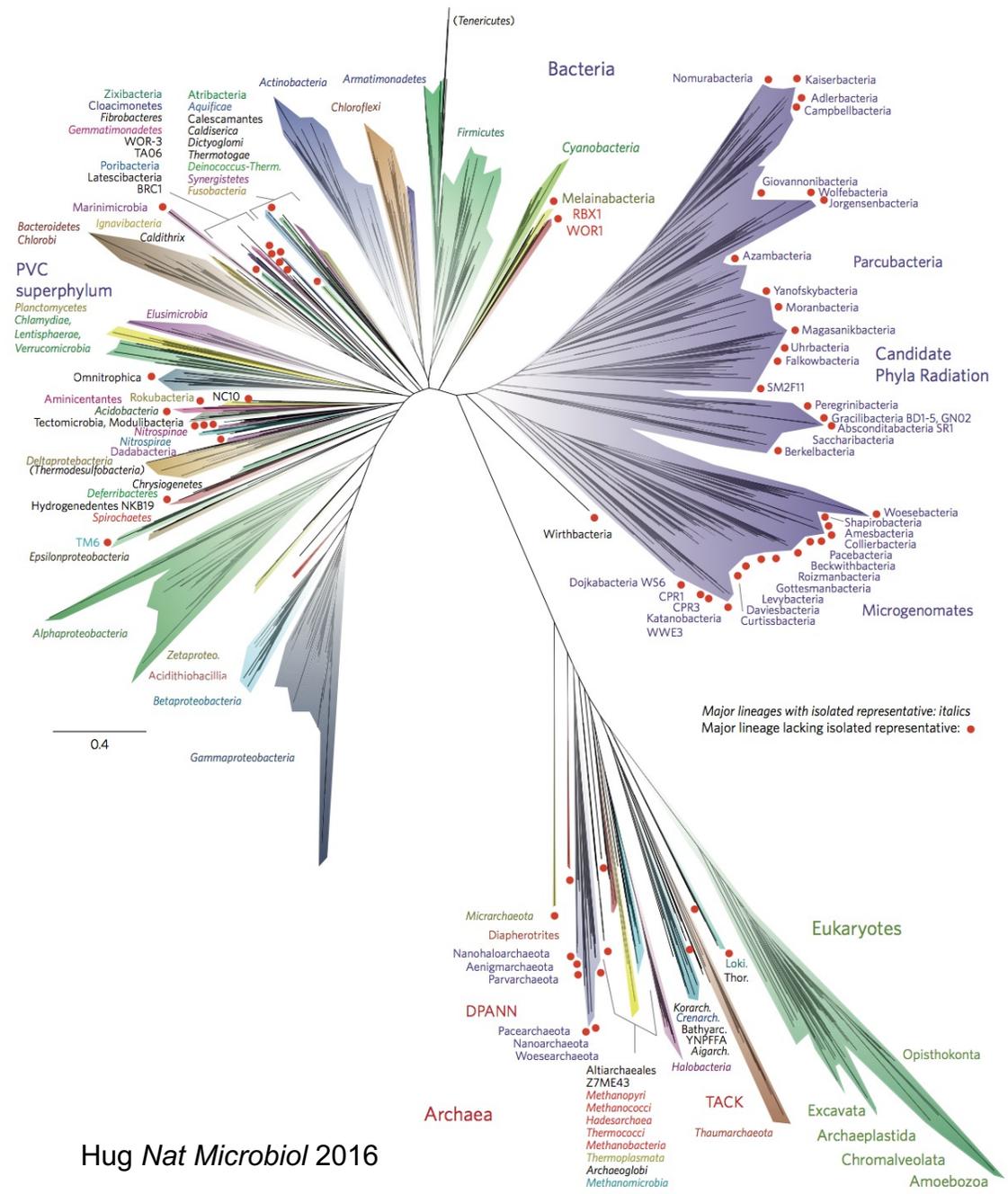
Generally considered too

variable, labour intensive, time-consuming, insufficiently discriminatory and poorly reproducible

Typing then moved into “molecular” or DNA-based era

Typing: why molecular?

- Genomes **define** the ancestry and relatedness of isolates
- Any way to access this information can provide better typing results



Molecular typing

Analyses discriminatory parts of the genome in the lab

- often amplification of specific regions / loci followed by separation on a gel and reading the pattern

Pulsed Field Gel Electrophoresis (PFGE)

Amplified fragment length polymorphisms (AFLP)

Random amplification of polymorphic DNA (RAPD)

Repetitive-element polymerase chain reaction (rep-PCR)

Variable-number tandem repeat (VNTR)

Staphylococcus aureus protein A gene-typing (spa)

Multilocus Enzyme Electrophoresis (MLEE – protein variant based)

Ribotyping (*Clostridioides difficile*)

Multi-locus sequence typing (MLST)

Different approaches are more suitable for different species

Molecular typing

Some techniques useful across the whole species

- And some offer higher resolution within closely related strains (“clones”)

Pulsed Field Gel Electrophoresis (PFGE)

Amplified fragment length polymorphisms (AFLP)

Random amplification of polymorphic DNA (RAPD)

Repetitive-element polymerase chain reaction (rep-PCR)

Variable-number tandem repeat (VNTR)

Staphylococcus aureus protein A gene-typing (spa)

Multilocus Enzyme Electrophoresis (MLEE – protein variant based)

Ribotyping (*Clostridioides difficile*)

Multi-locus sequence typing (MLST)

Because they are based on different genomic features

Pulsed Field Gel Electrophoresis

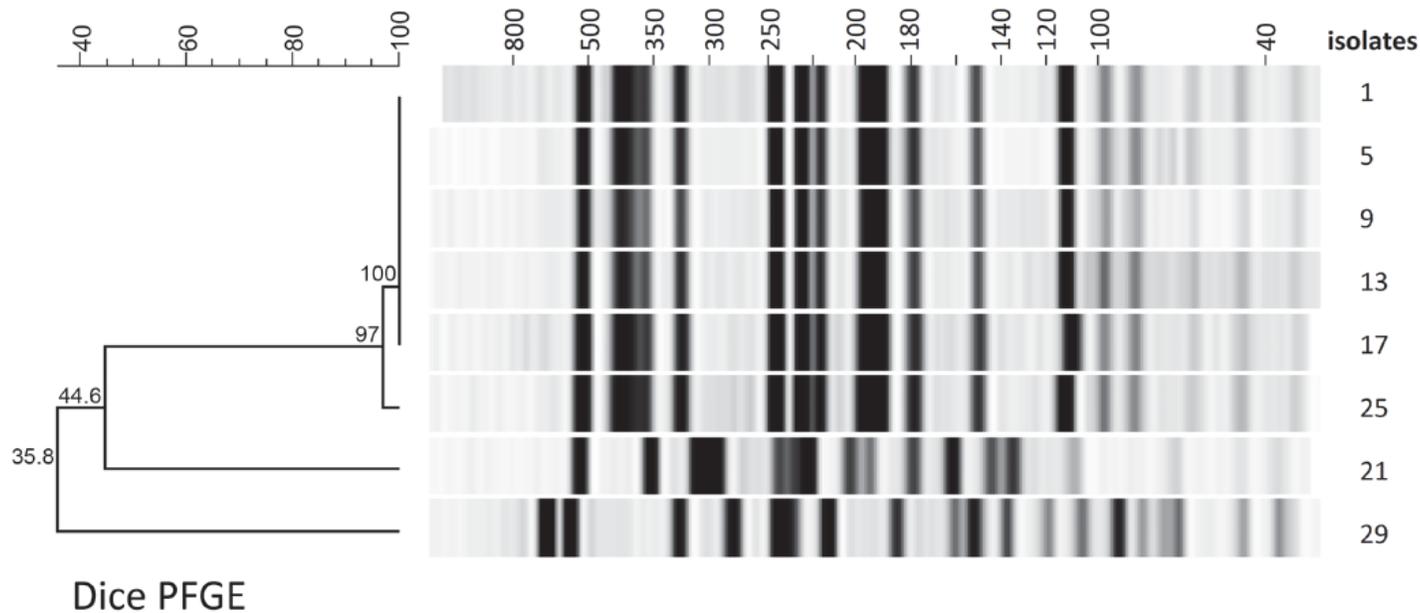
Was reference standard for many years

Comparison of band patterns

Algorithm determines the phylogeny

Few databases available

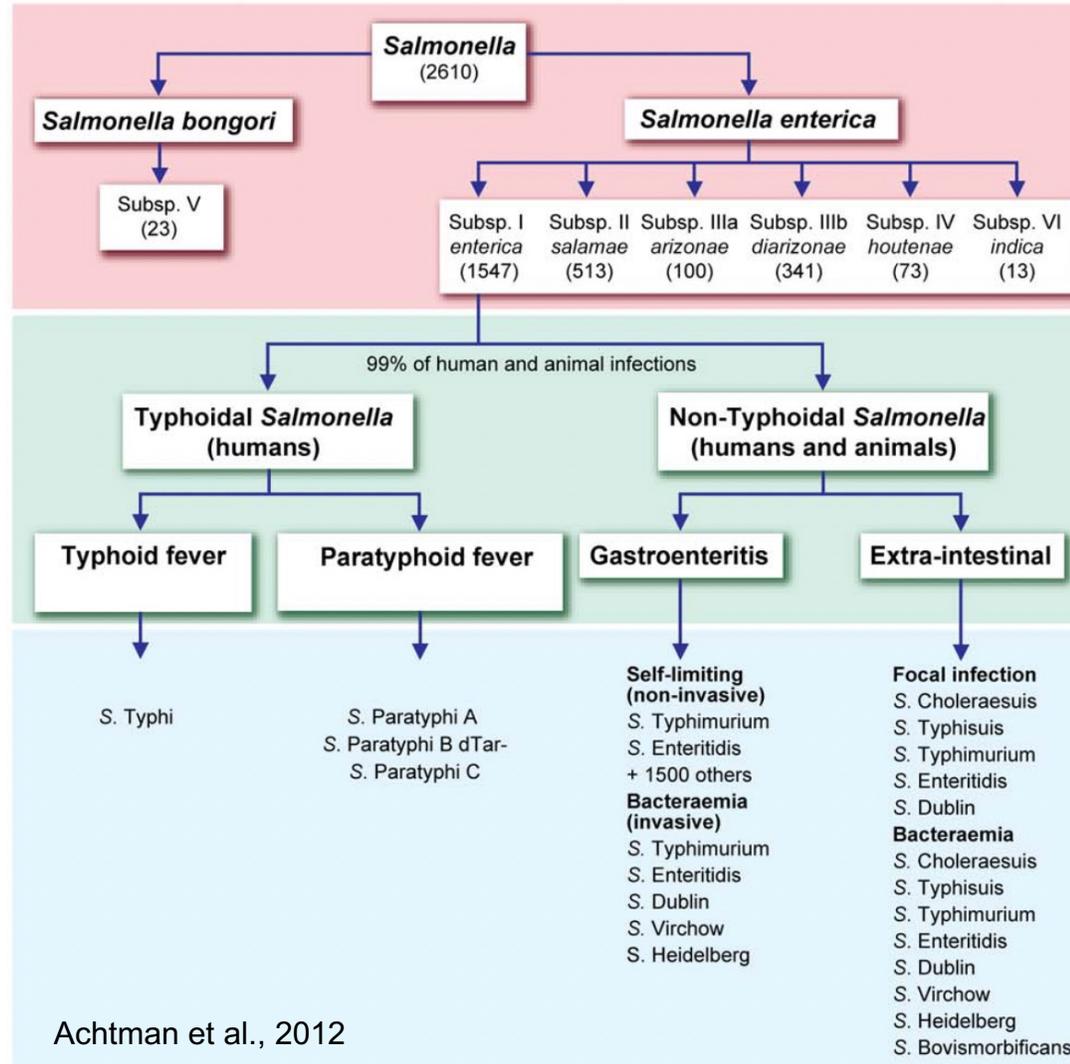
Low reproducibility between centers



E. coli PFGE
 Outbreak and two unrelated isolates
 Egli *PLoS ONE* 2015

Salmonella serotyping (and more)

- *Salmonella* Typhi (*Salmonella enterica* subspecies *enterica* serotype Typhi) is a serovar of a disease-syndrome defined group of a subspecies
- *Salmonella* serotyping depends on specific agglutination reactions with adsorbed antisera specific for O or H antigens
- Antigen + appropriate antibody = agglutination



Species and subspecies were originally defined by DNA-DNA hybridisation, confirmed by MLEE and MLST and are currently differentiated by biochemistry and serology.

The split in typhoidal and non-typhoidal is based on the disease syndrome. Typhoid and paratyphoid fever is prolonged, whilst extra-intestinal infection is usually acute and metastatic. Gastroenteritis is characterised by diarrhoea.

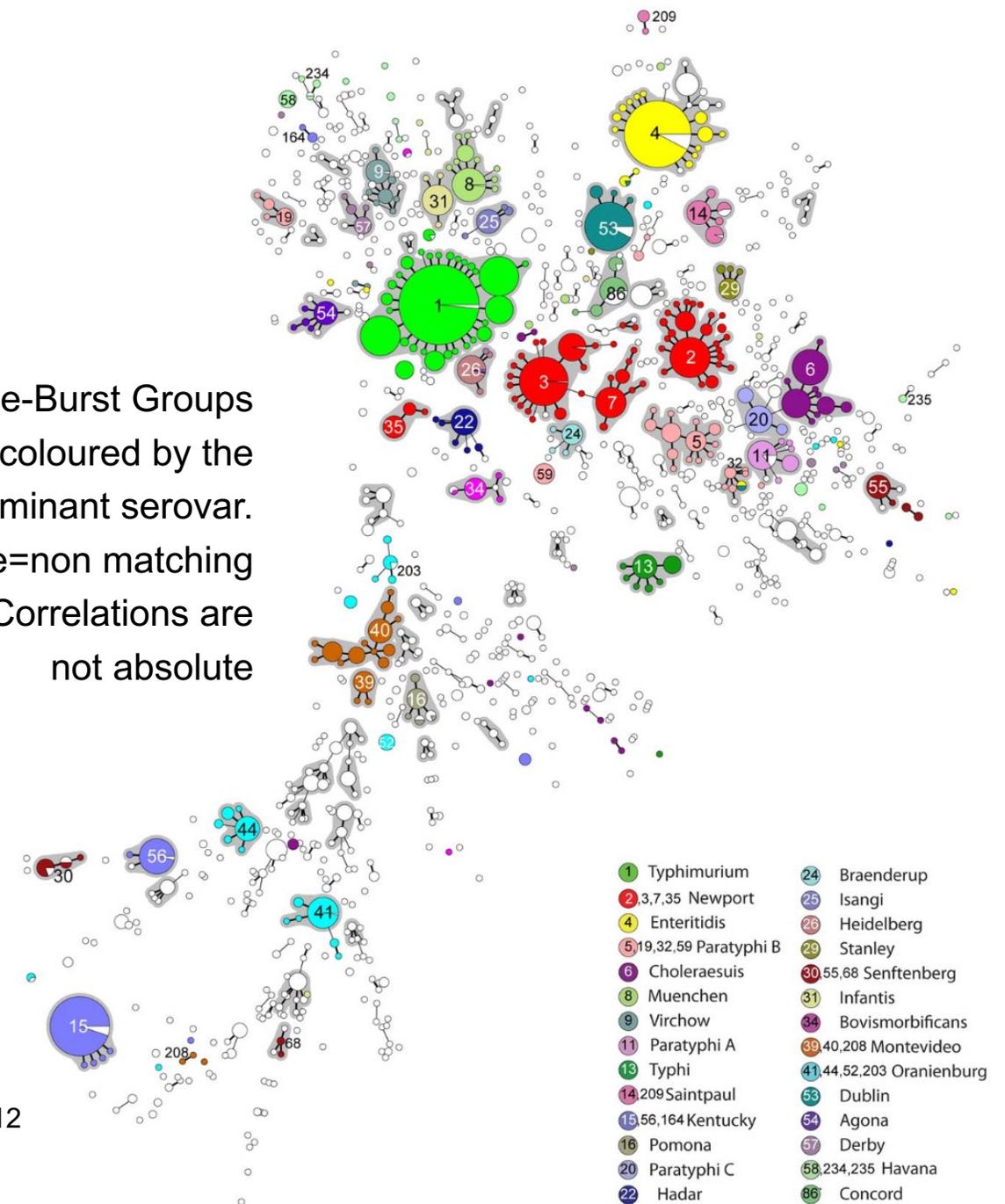
Differentiation of serovars is by agglutination with specific antisera against LPS (O), two phases of flagella (H1 and H2). There are 46 O, 85 H and 1 capsule (Vi) antigen which have been described in about 1,500 combinations within subspecies I.

Figure 1. General overview of the current classification of *Salmonella enterica*. doi:10.1371/journal.ppat.1002776.g001

Salmonella MLST

- *Salmonella* Typhi (*Salmonella enterica* subspecies *enterica* serotype Typhi) is a serovar of a disease-syndrome defined group of a subspecies
- *Salmonella* serotyping depends on specific agglutination reactions with adsorbed antisera specific for O or H antigens
- Comparison with MLST (7 gene fragments): “serovar designations confounded genetically unrelated isolates and failed to recognize natural evolutionary groupings”

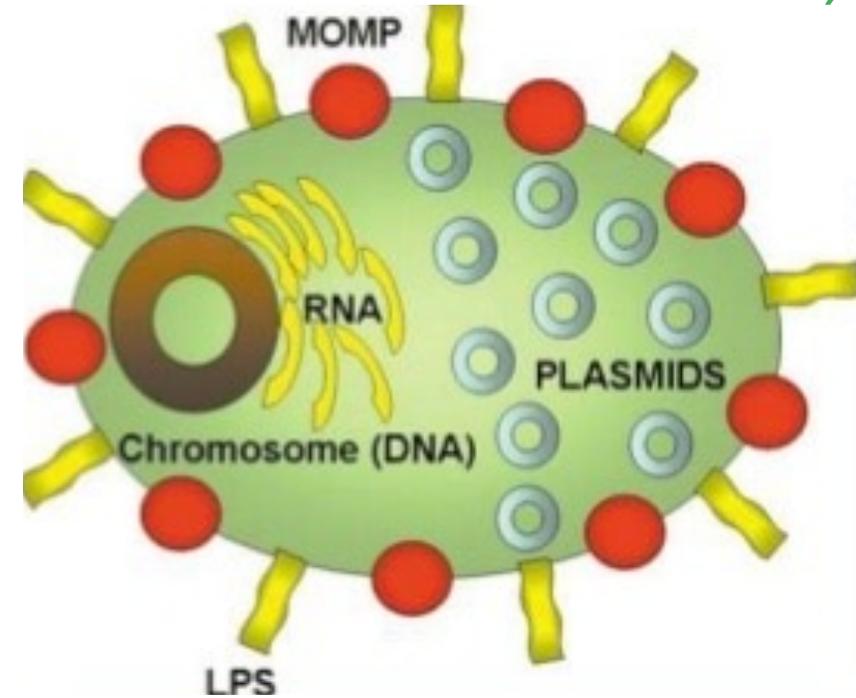
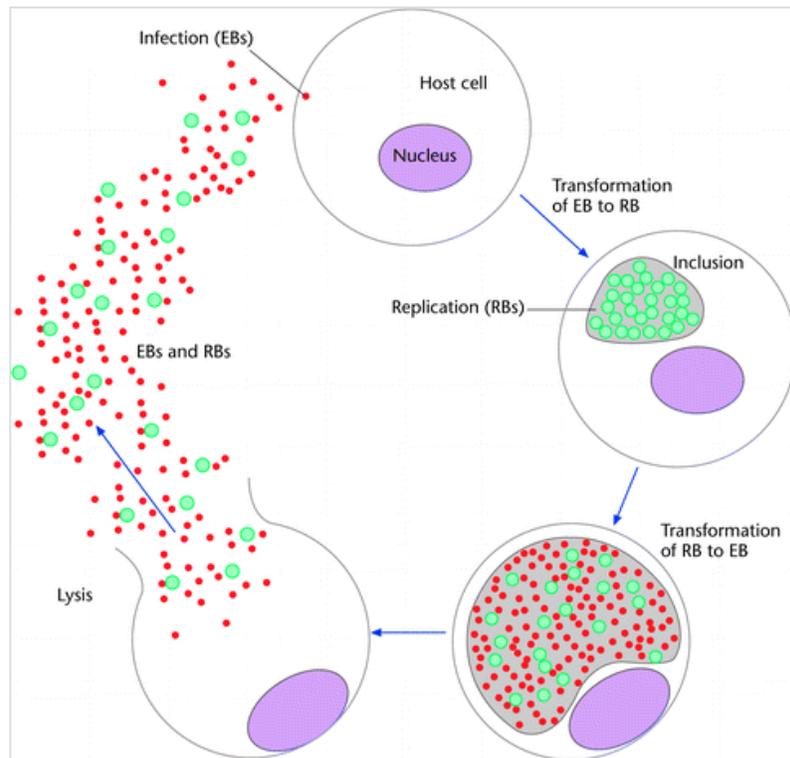
Groups / e-Burst Groups are coloured by the dominant serovar. White=non matching serovar. Correlations are not absolute



Achtman et al., 2012

C. trachomatis: serotyping became genotyping

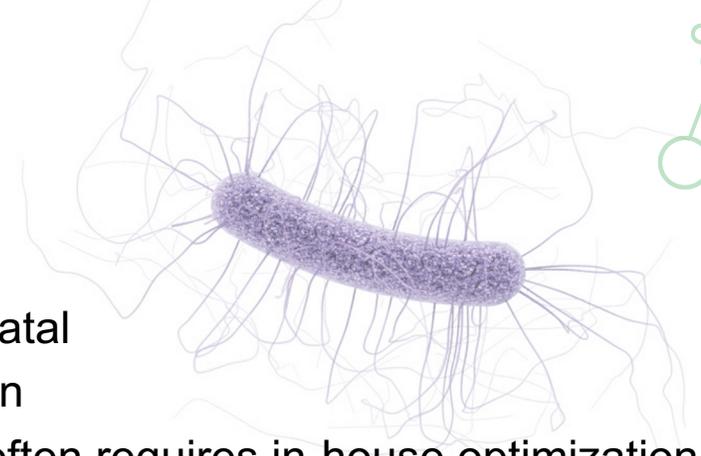
- Gram negative
- Intracellular
- Small genome ~1 Mb
- Few gene differences across species



- MOMP: 60% of outer membrane protein mass
- serotyping with monoclonal antibodies from eg. HeLa 229 cells
- MOMP encoded by gene *ompA* -> genotyping

Stephens *Science* 1998 (Trachoma); Carlson *Inl* 2005 (STI / Urogenital); Thomson *Genome Res* 2008 (LGV); Seth-Smith *BMC Genomics* 2009

Clostridioides difficile: ribotyping



- Important nosocomial pathogen
- Asymptomatic -> *C. difficile* infection (CDI) -> severe colitis, sepsis, fatal
- PCR-ribotyping commonly used typing tool: good strain discrimination
- BUT not fully portable between laboratories, labour intensive, slow, often requires in-house optimization
- Hypervirulent lineages have been defined by ribotype: RT027 and RT078
- What is the connection between ribotype, virulence and genome?

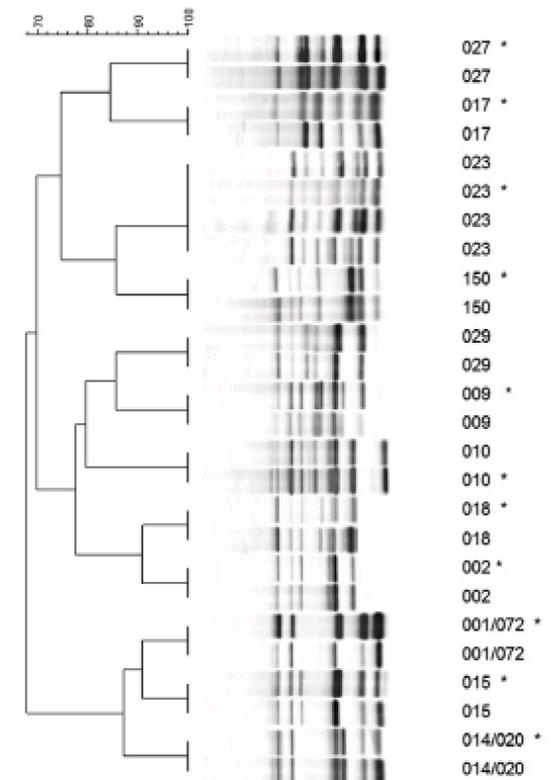
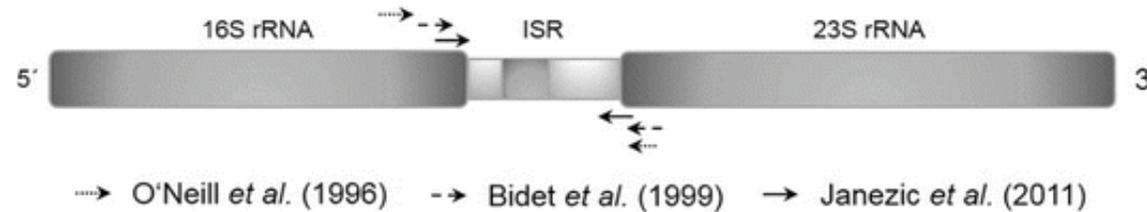
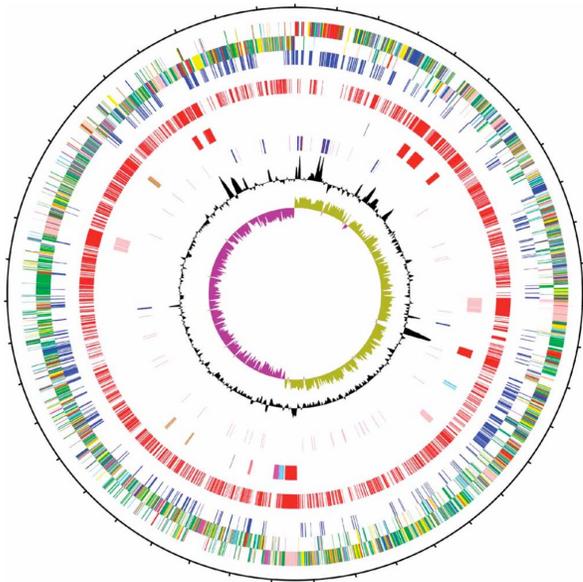
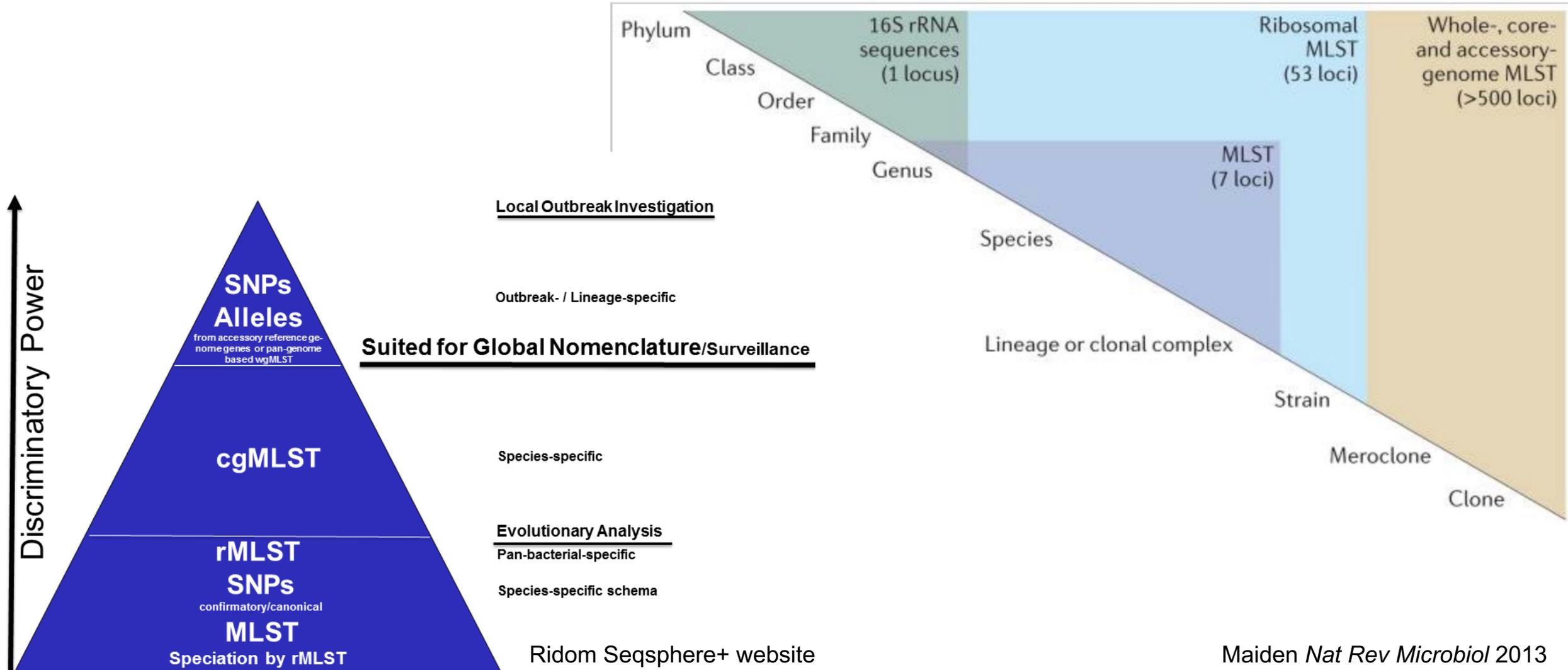


Fig. 2 Comparison of PCR-ribotyping patterns obtained from total stool DNA (marked with *) and reference strains using primers and protocol described in Janezic et al. [20] and in this chapter

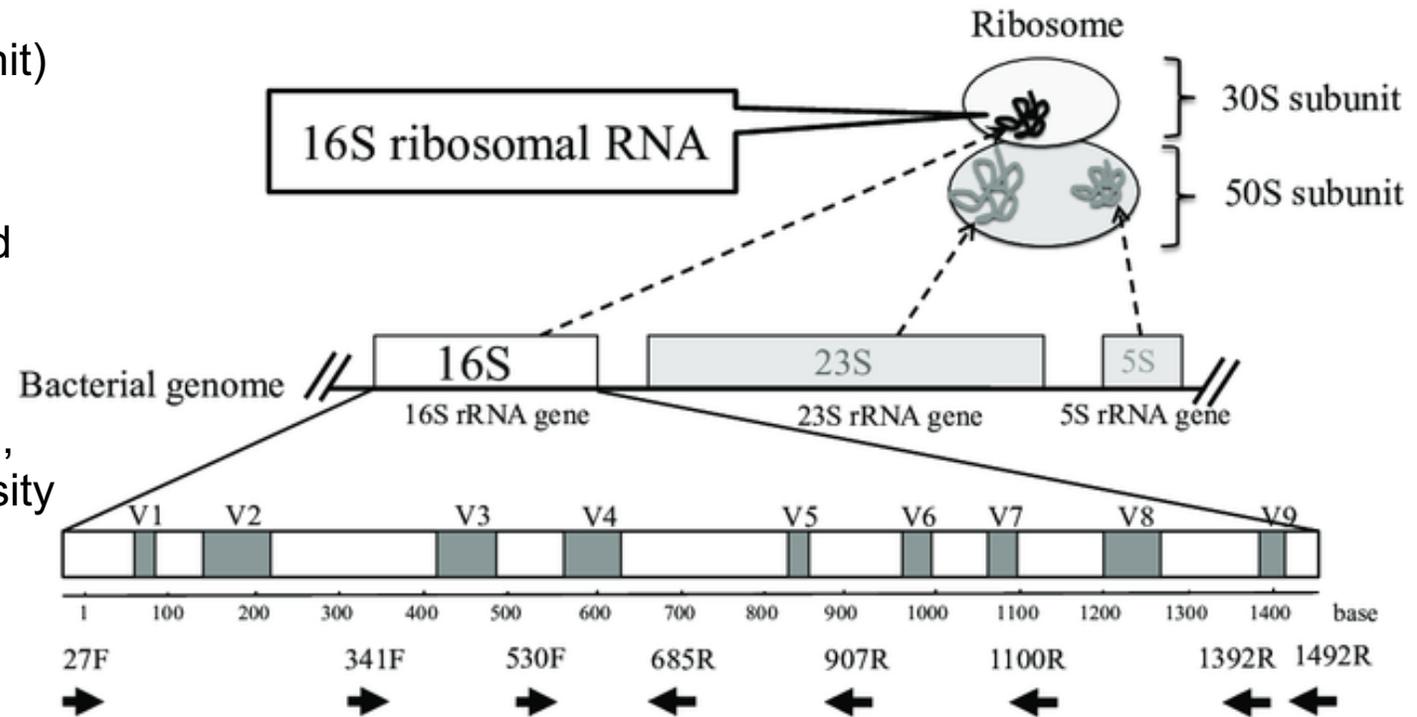
Sebahia *Nat Gen* 2006; Janezic *Clostridium difficile: Methods and Protocols* 2016; *C. difficile* Methods and Protocols, Springer

Genome-based typing methods: resolution



16S ribosomal RNA typing

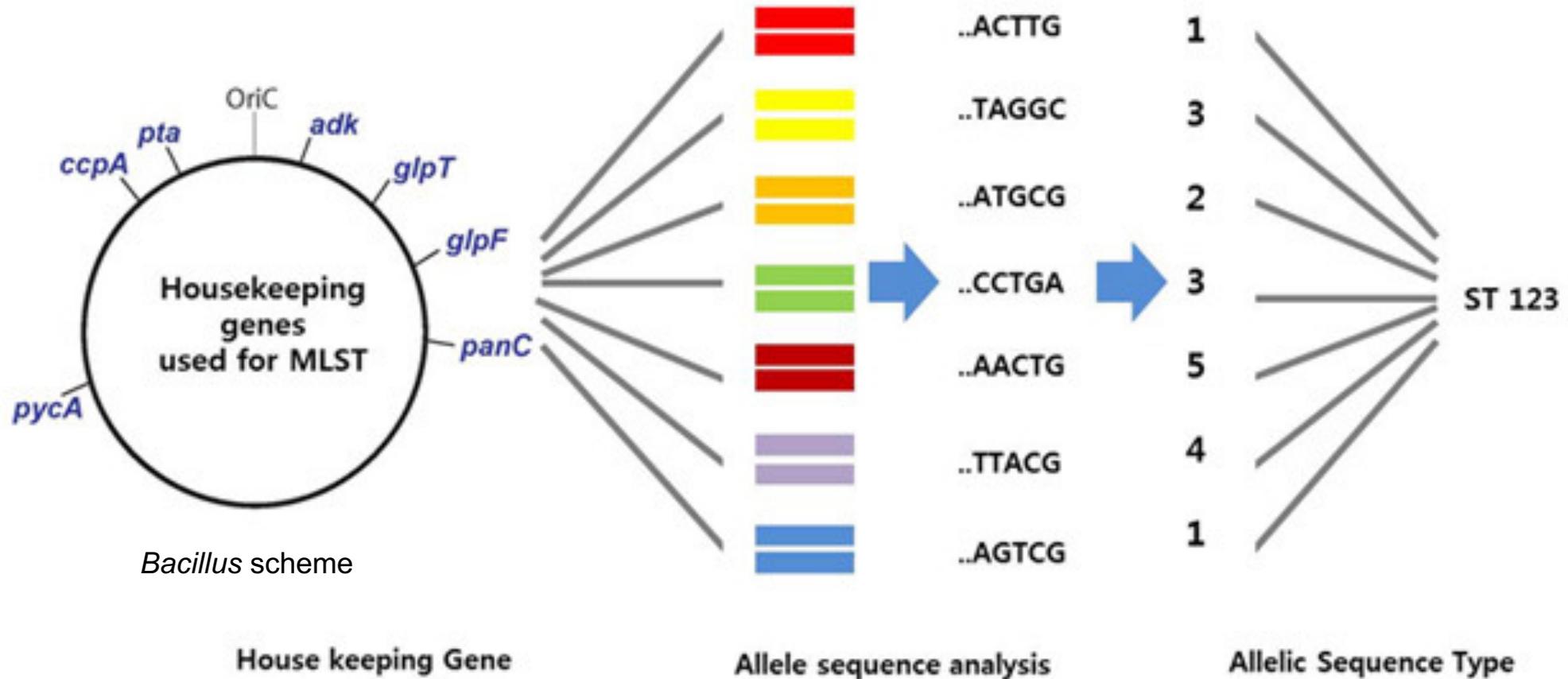
- 16S rRNA gene produces 16S rRNA
- Forms part of ribosome (part of 30S subunit)
- Sufficiently conserved / diverse to define species when sequenced
- V / variable regions are frequently targeted
- Species correlation not always perfect
- Some genomes have several copies: 2-14, average 6, and they can have some diversity



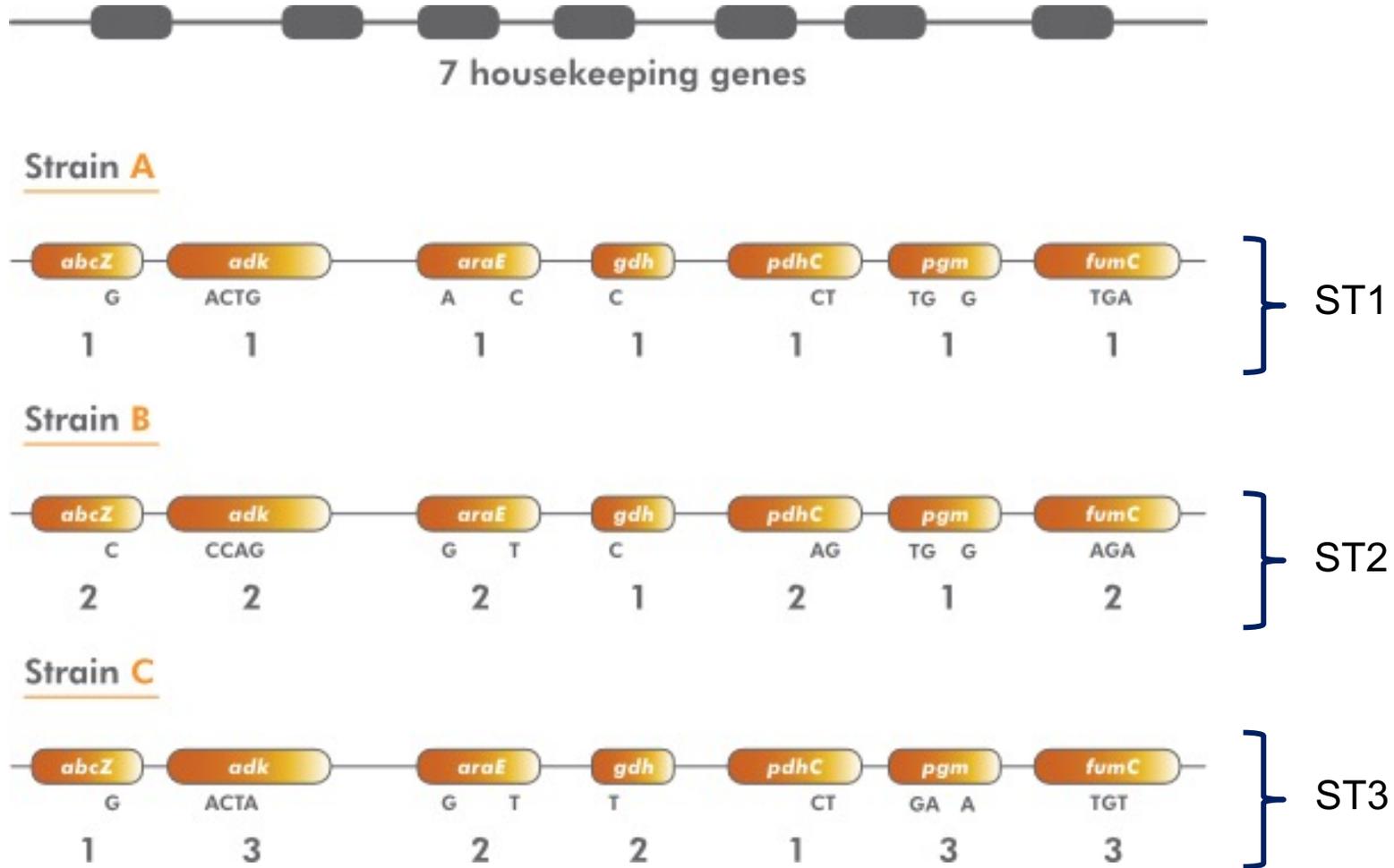
DOI: 10.7888/juoeh.38.223

Typing methods: MLST

- Manageable number (5-7) of conserved (housekeeping) genes to represent genome diversity
- Originally by PCR and Sanger sequencing: data can also be extracted from WGS
- First in Maiden *PNAS* 1998; *Neisseria meningitidis*; 6 loci chosen to reflect resolution of MLEE



How does MLST work?



Biomérieux, Applied Maths, Bionumerics website

PubMLST Public databases for molecular typing and microbial genome diversity Disconnect MY ACCOUNT

[HOME](#) [ORGANISMS](#) [SPECIES ID](#) [ABOUT US](#) [UPDATES](#)

A collection of open-access, curated databases that integrate population sequence data with provenance and phenotype information for over 130 different microbial species and genera.

33,973,366 ALLELES 1,038,001 ISOLATES 768,736 GENOMES



- Schemes defined for many species: international, sharable...

Organisms search APPLY

Organisms
Choose your organism from a list of over 130 species and genera-specific databases. Access molecular typing and isolate records.

Species ID
Use ribosomal MLST to accurately identify bacterial species from a genome assembly.

Global Meningitis Genome Library

Sexually transmitted infections

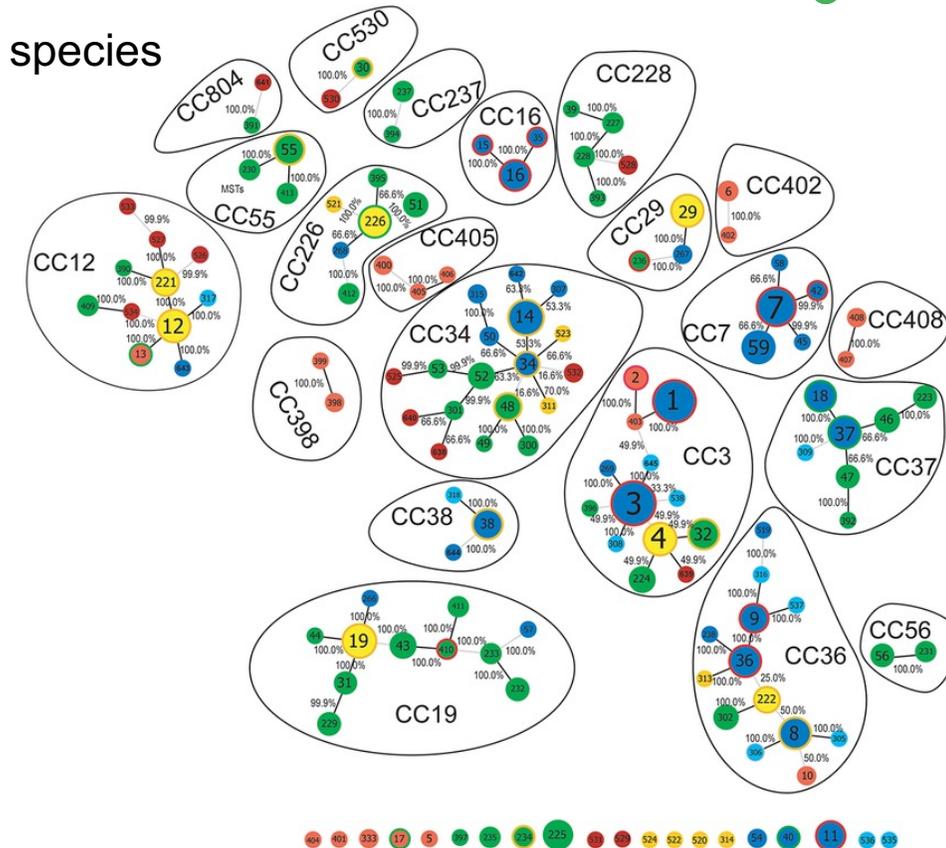
Designed to be portable “across the World-Wide Web site, thus enabling exchange of molecular typing data for global epidemiology via the Internet.”
Maiden *PNAS* 1998

Clonal Complexes

Clonal complexes are groupings of MLST sequence types, in some species

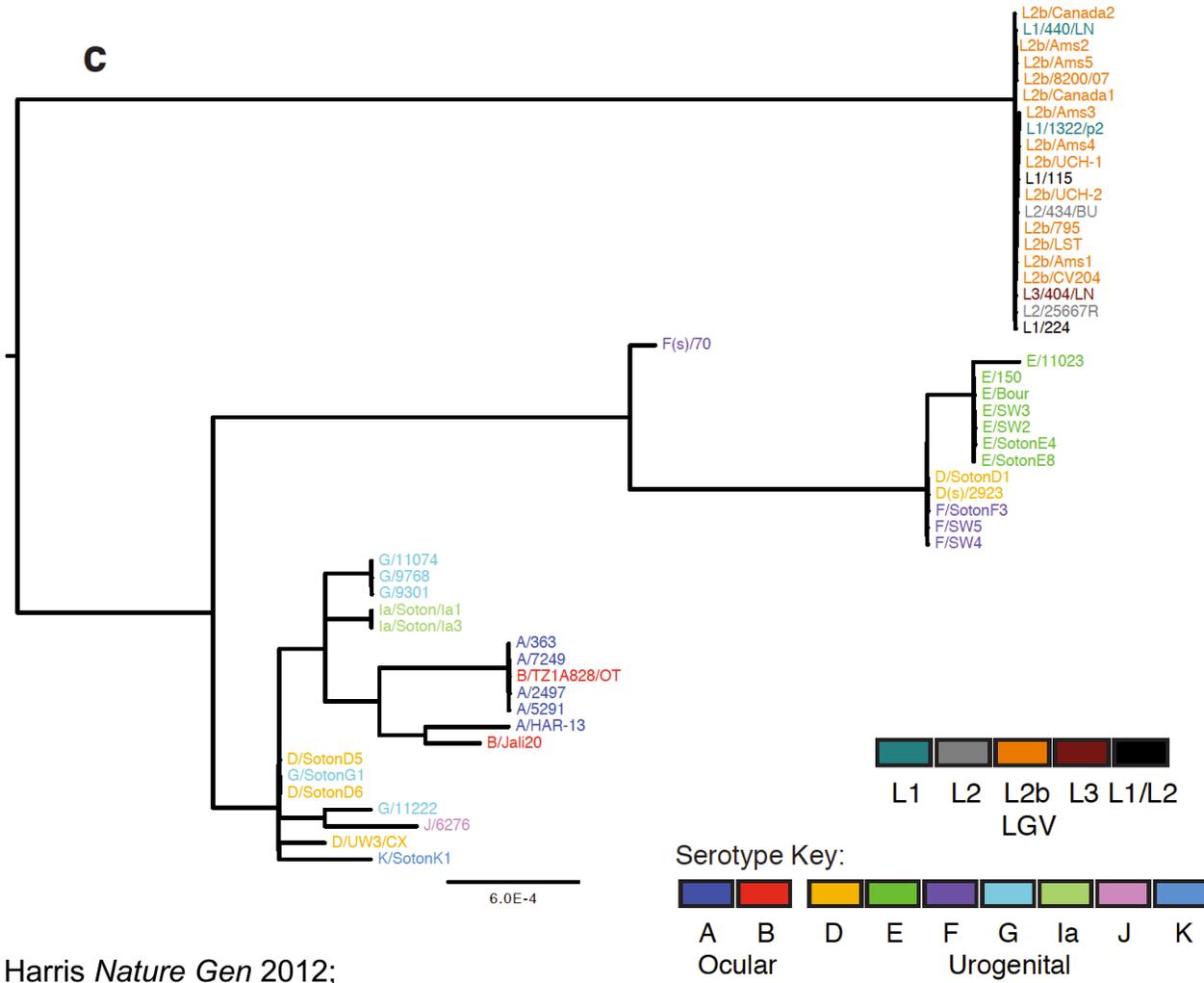
Grouped according to a specific number of shared alleles, eg:

- *Staphylococcus aureus*: 4+ shared loci
- *Borrelia burgdorferi*: single / double locus variants (figure)



10.1371/journal.pone.0149345

MLST representation: SNP tree of concatenated loci



Harris *Nature Gen* 2012;
Chlamydia trachomatis

Neisseria meningitidis: hypervirulent lineages and MLST

- Most important hyper-virulent lineages are subgroups I, III, and IV-Other, by MLEE
- Comparison of MLST results:
 - serogroup A strains formed a distinct cluster of lineages
 - major subgroups associated with epidemic meningitis (I, III, and IV-1) were easily distinguished

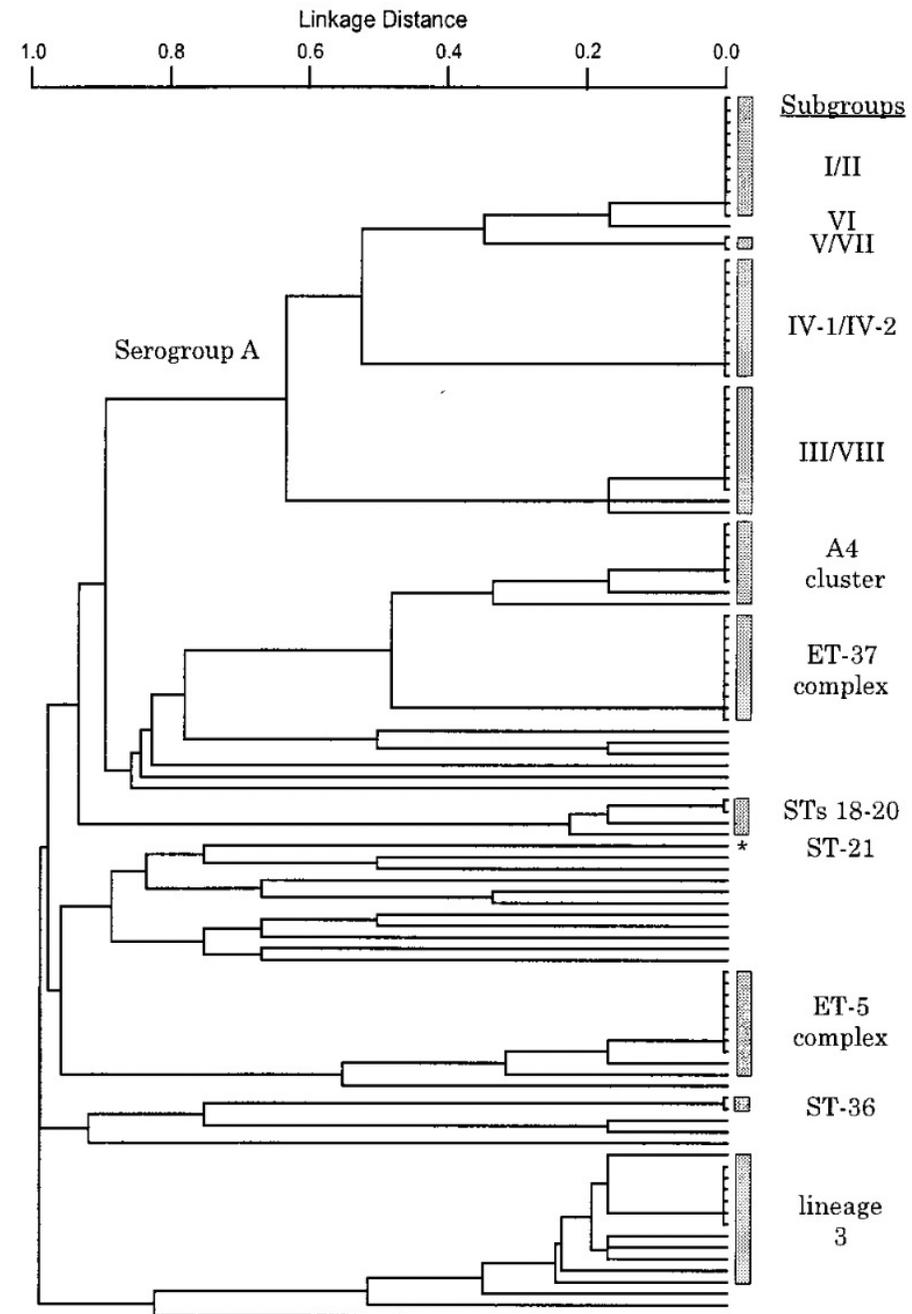
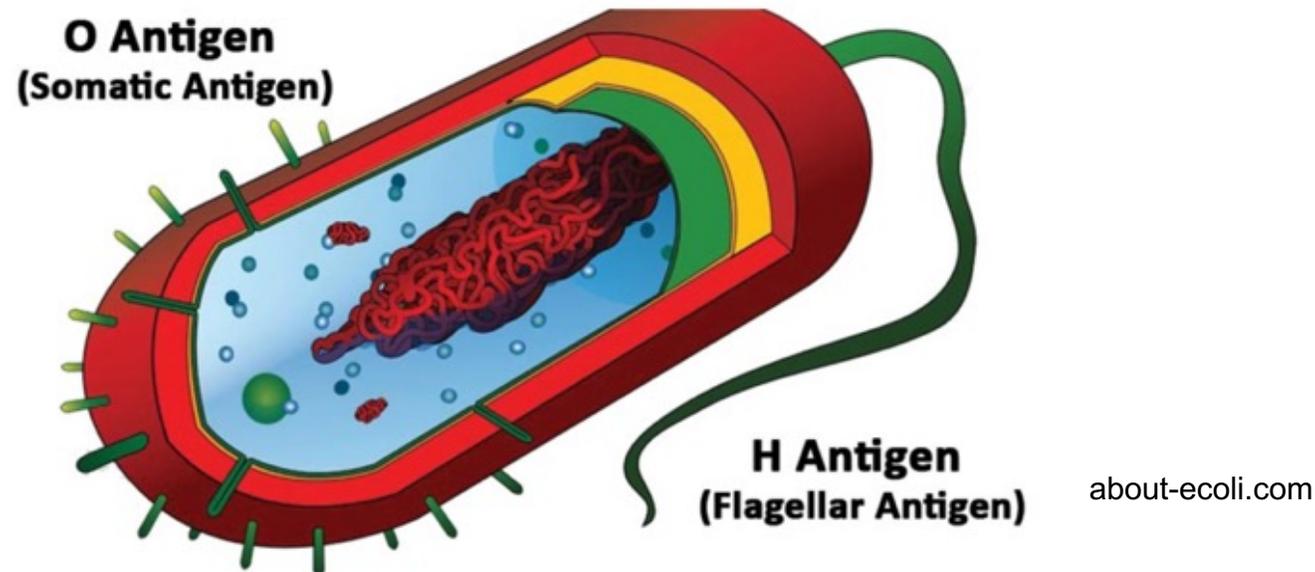


FIG. 2. Dendrogram of genetic relationships among 107 strains

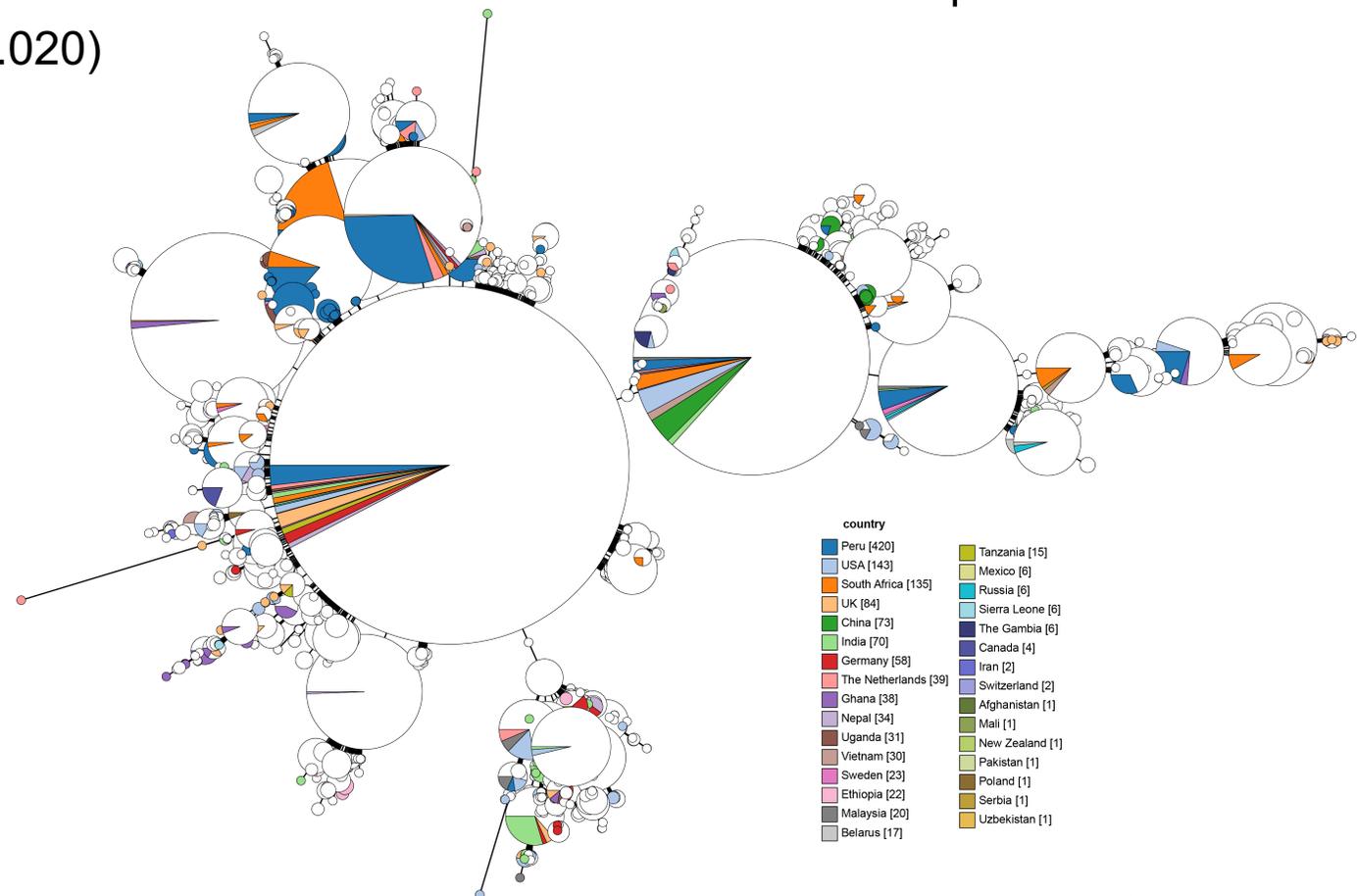
E. coli typing – disease association

- *E. coli* O157:H7 commonly associated with disease in humans
 - Based on O and H antigen serotyping
 - Also Shiga toxin typing based on serology / amino acid sequence predicted from genome
- Different molecular techniques give different resolutions:
 - Randomly amplified polymorphic DNA (RAPD) and pulsed-field gel electrophoresis (PFGE) given resolution within O157:H7. Multi-locus sequence typing (MLST) doesn't. Typing using techniques using whole genome data provide highest resolution. (doi.org/10.1038/s41538-021-00097-0)

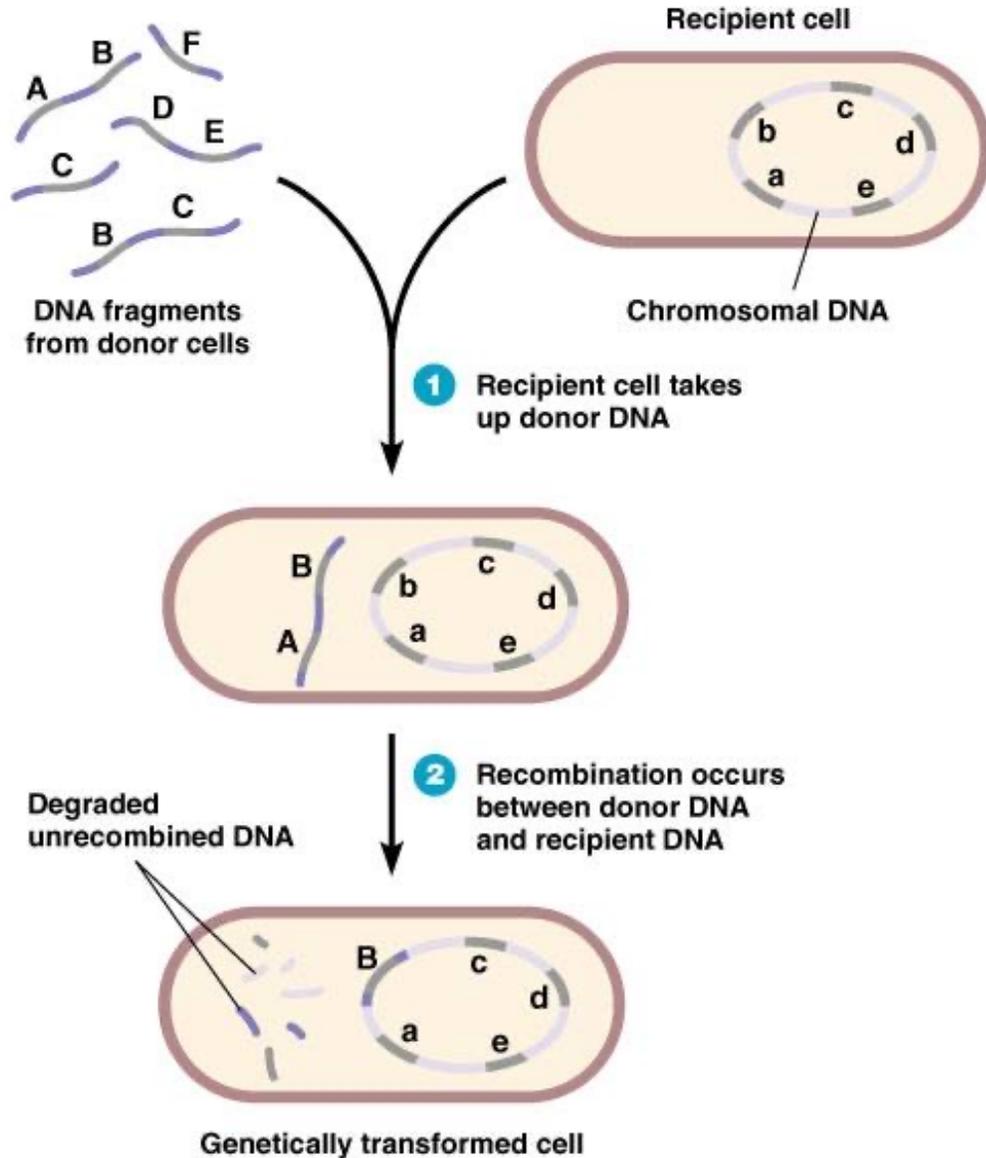


Mycobacterium MLST

- Scheme devised for *Mycobacterium* genus
- Success among non-tuberculous Mycobacteria (NTMs)
- Limited genomic diversity in *Mycobacterium tuberculosis* means it is not a useful technique in this species (doi.org/10.1016/j.mimet.2013.01.020)
- Figure generated from all *Mycobacterium tuberculosis* data in PubMLST



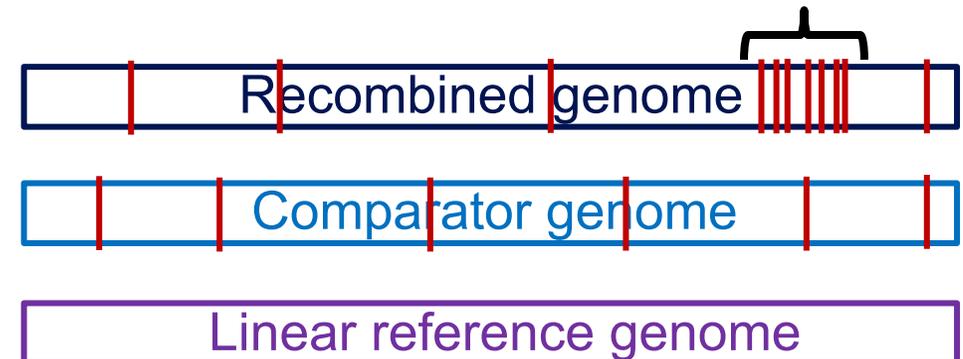
Recombination can confound typing



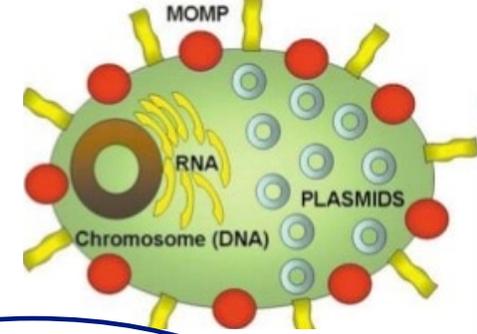
How to identify this is genome sequence:

Mutations (SNPs) often accumulate randomly

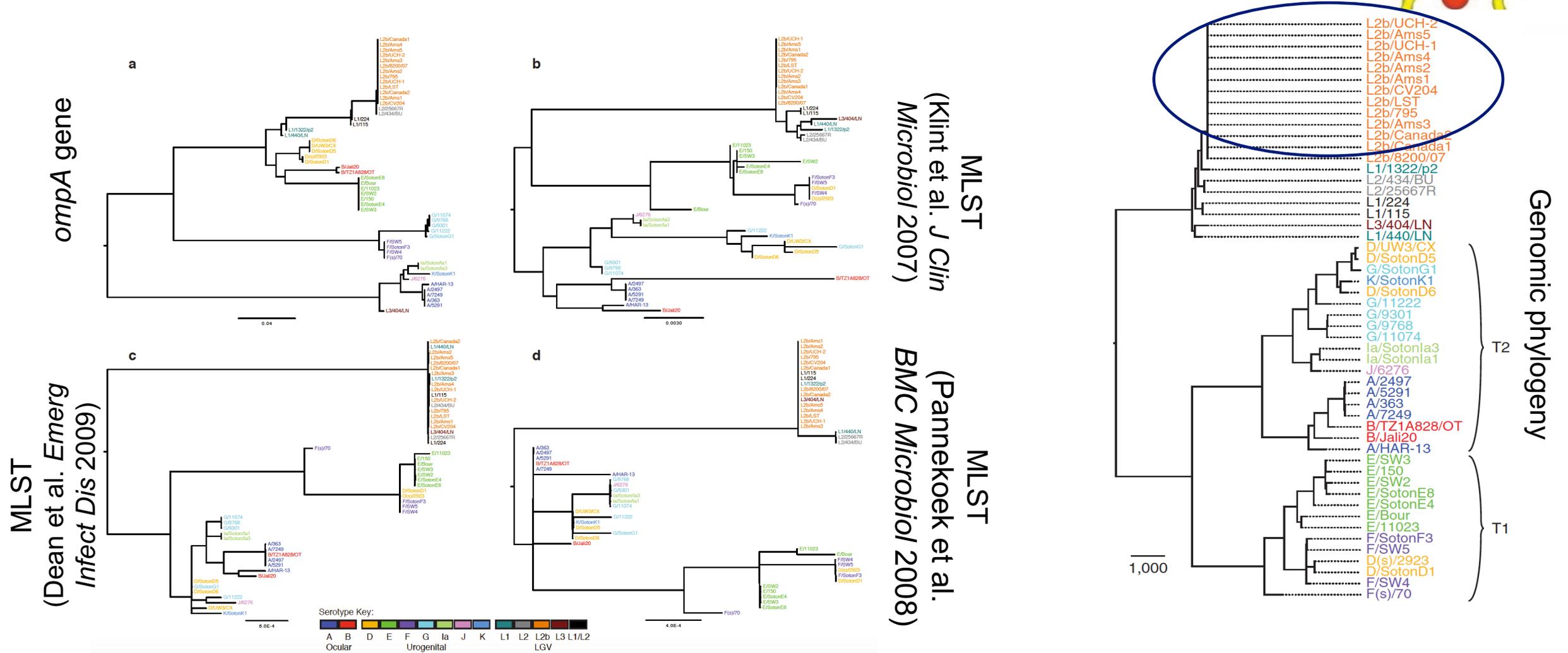
High density of SNPs implies gain of "foreign" DNA



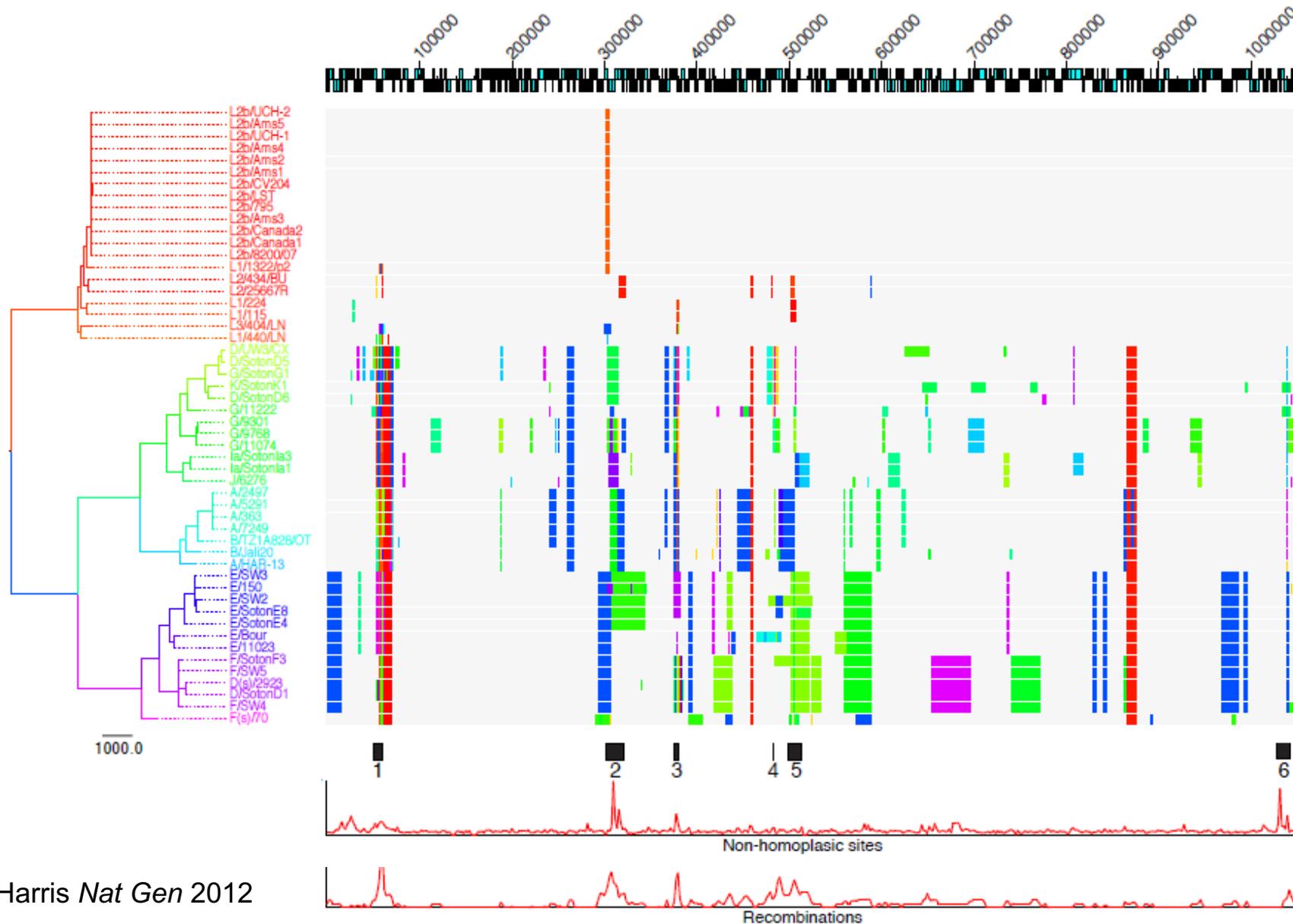
Recombination can confound typing: *Chlamydia trachomatis*



Colour mixing (tip name labels) shows how MOMP / *ompA* typing disagrees with full genome data
 Tree structures show how resolution differs



Recombination in *C. trachomatis*

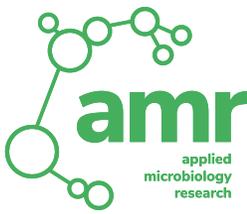


Large scale recombination:

- 26% of sites (4492 /17163) are homoplastic

Certain loci are more variable than others

Discussion



- Conventional typing was based on phenotypes
- Almost all typing now is based on DNA analysis
- The more information included, the more accurate
- MLST is based on a few conserved loci per species: can be analysed by PCR and Sanger sequencing or from the whole genome
- The nomenclatures behind typing are pervasive, especially among clinicians

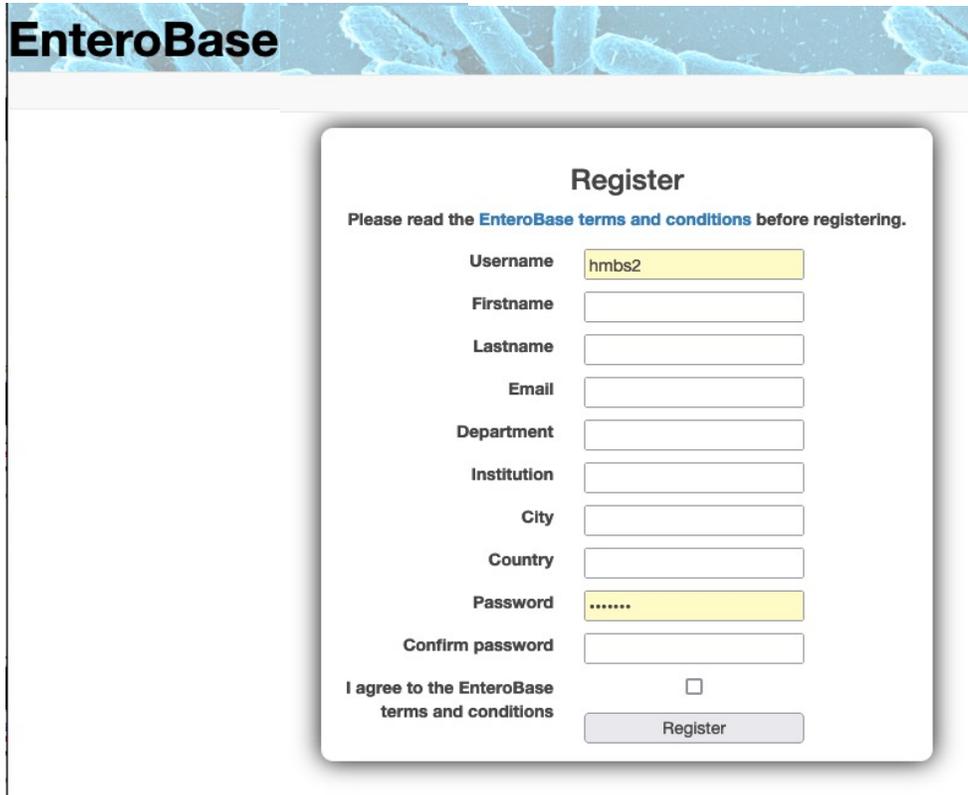
- Stay tuned for more information on genome-based typing tomorrow!

Small exercise

Please generate logins, for tomorrow's work, at:

Enterobase : <https://enterobase.warwick.ac.uk/auth/register>

PathogenWatch: <https://pathogen.watch/sign-in>



Enterobase

Register

Please read the [Enterobase terms and conditions](#) before registering.

Username

Firstname

Lastname

Email

Department

Institution

City

Country

Password

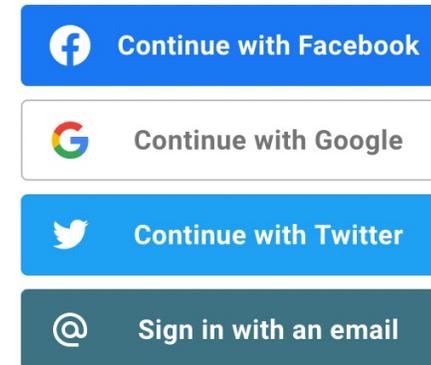
Confirm password

I agree to the Enterobase terms and conditions



☰  Pathogenwatch GENOMES COLLECTIONS UPLOAD DOCUMENTATION   

Sign in to your Pathogenwatch account



 Continue with Facebook

 Continue with Google

 Continue with Twitter

 Sign in with an email

Reading suggestions

Introduction to microbial bioinformatics

<https://www.sciencedirect.com/science/article/pii/S198743X17307097?via%3Dihub>

Recommended review

Balloux: From Theory to Practice: Translating Whole-Genome Sequencing (WGS) into the Clinic, 10.1016/j.tim.2018.08.004

rMLST:

<https://doi.org/10.1099/mic.0.055459-0>

PubMLST and BIGSdb:

<https://wellcomeopenresearch.org/articles/3-124/v1>

Enterobase

<https://genome.cshlp.org/content/30/1/138.long>

**Many thanks for your
attention**

Questions??